

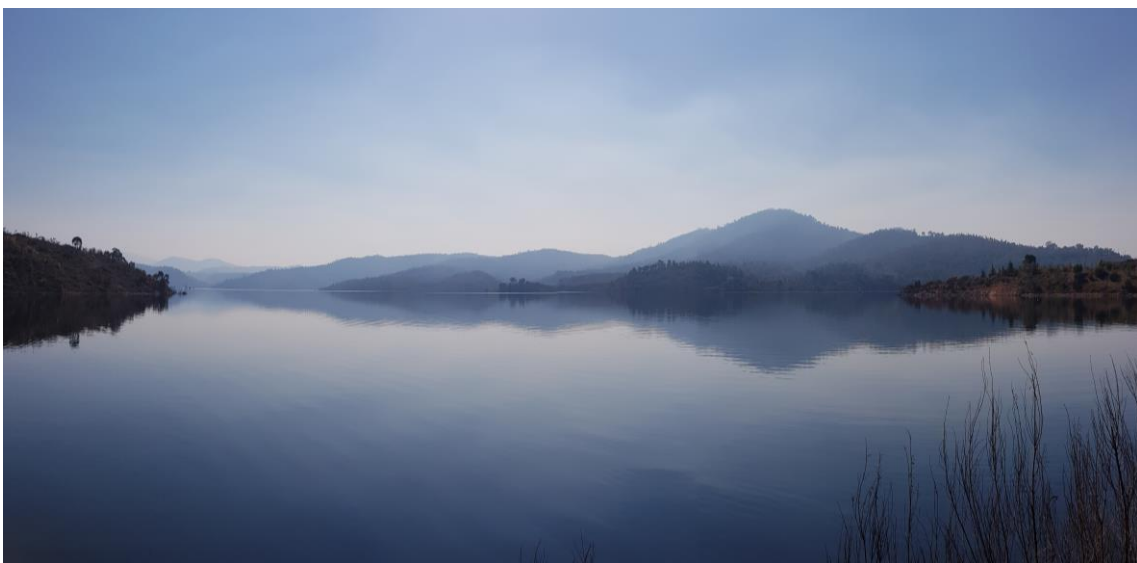


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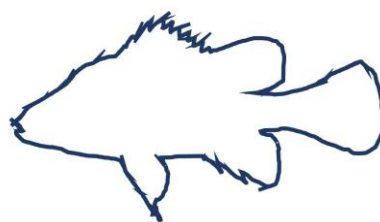
Enlarged Cotter Reservoir Ecological Monitoring Program

Technical Report 2022



Report to Icon Water

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Enlarged Cotter Reservoir Ecological Monitoring Program: Technical Report 2022

Report prepared for: Icon Water

Authors: Ben Broadhurst, Rhian Clear, Hugh Allan and Mark Lintermans

Produced by:

Centre for Applied Water Science

Institute for Applied Ecology

University of Canberra, ACT 2601

Telephone: (02) 6206 8608

Facsimile: (02) 6201 5651

Website: <http://www.canberra.edu.au/centres/iae/index.php>

ABN: 81 633 873 422

Inquiries regarding this document should be addressed to:

Ben Broadhurst

Phone: 0423 363 636

Fax: (02) 6201 5305

Email: ben.broadhurst@canberra.edu.au

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EXECUTIVE SUMMARY

Ongoing drought and its threat to water security in the ACT resulted in the recommissioning and augmentation of Cotter Reservoir from ~4 GL to 76.2 GL capacity, known as the Enlarged Cotter Reservoir (ECR). The ECR and Cotter River upstream to Bendora Dam contain four threatened fish and crayfish species, though only Macquarie perch *Macquaria australasica* and Two-spined blackfish *Gadopsis bispinosa* are likely to be directly impacted by the ECR and consequently are the focal species for research and mitigation projects associated with the ECR. Potential impacts of the construction, filling and operation of ECR have been well-described and in response to these impacts a range of projects including this fish monitoring program have been undertaken. A monitoring program commenced in 2010 with baseline monitoring (pre-filling) completed in 2013.

Since 2013, this ecological monitoring program centres on 10 management questions that aim to determine the impact of the filling and operation of the ECR on populations of the two focal species and potential threats (predators and competitors) in the ECR and river upstream. The post-2013 monitoring encompasses the ECR filling phase, since filling commenced in April 2013. This report addresses the 10 management questions by comparing baseline (2010 – 2013, (see Lintermans *et al.* 2013), and filling (2014 and 2015) (Broadhurst *et al.* 2014, Broadhurst *et al.* 2015) and operational (2016 – 2022)(Broadhurst *et al.* 2016b, Broadhurst *et al.* 2017, Broadhurst *et al.* 2018, 2019, Broadhurst *et al.* 2020, 2021) monitoring data (where possible).

The 10 management questions that underpin the Enlarged Cotter Reservoir Ecological Monitoring Program since 2013 are:

1. Has there been a significant change in the abundance and body condition of Macquarie perch in the enlarged Cotter Reservoir (Young-of-Year, juveniles and adults) as a result of filling and operation?
2. Has there been a significant change in the abundance, body condition and distribution of the Macquarie perch in the Cotter River above and below Vanities Crossing as a result of the filling and operation of the ECR?
3. Have Two-spined blackfish established a reproducing population in the enlarged Cotter Reservoir and are they persisting in the newly inundated section of the Cotter River?
4. Has there been a significant change in the abundance, distribution and size composition of adult trout in the enlarged Cotter Reservoir as a result of filling and operation?
5. Has there been a significant change in the abundance and size composition of trout in the Cotter River upstream of the enlarged Cotter Reservoir as a result of the filling and operation?
6. Are Two-spined blackfish and Macquarie perch present in trout stomachs in the Cotter River?
7. Has there been a significant change in the abundance and distribution of non-native fish species) in the enlarged Cotter Reservoir as a result of the filling and operation?
8. Has there been a significant change in the abundance, distribution and species composition of piscivorous birds in the vicinity of the enlarged Cotter Reservoir as a result of the filling and operation?
9. Have macrophyte beds re-established in the enlarged Cotter Reservoir?
10. Are there adequate food resources (particularly decapods) for the Macquarie perch following the filling and operation of the enlarged Cotter Reservoir?

The monitoring year of 2021 / 2022 was relatively wet and Cotter River flows below Bendora were largely unregulated. There were several peaks in flow associated with rainfall events, most notably between November 2021 – January 2022. The Enlarged Cotter Reservoir (ECR) reached full supply

level in August 2020 and has remained functionally full (within 1.0 m of FSL) since. The ECR has now been in the 'operational' phase (i.e. it has filled and is now fluctuating in level with changing inflows and river management) since 2016. The unregulated elevated flows of the Cotter River prevented autumn sampling from being undertaken in 2022, which mean that questions 2, 5 and 6 could not be addressed for this monitoring year.

The main changes detected in the population of Macquarie perch in the ECR between the different monitoring phases (baseline, filling, operational) relate to adult abundance and body condition, and abundance of young-of-year recruits. Since peak abundances in 2015, adult relative abundance was in decline to its lowest level in 2018, whilst adult body lengths have been increasing. It appears that at least some of this trend may be due to changes in capture efficiency across size classes of Macquarie perch with changes in the ECR phases, with the gill nets deployed more effectively sampling smaller adults. Adult numbers as determined by gill netting in 2022 were low, though captures of adult Macquarie perch via boat electrofishing was very high compared to other years. This indicates that the relative abundance of adult Macquarie perch in Cotter Reservoir may not have reduced, but the lower captures may be because of a change in capture efficiency of gill nets. It is possible that a change in behaviour of adult Macquarie perch has rendered them less likely to be captured by gill nets in 2022. Body condition of adults was higher during filling and early operational phases, compared to baseline. Encouragingly, successful recruitment to young-of-year stage was detected for the fifth consecutive year in 2022, which is a positive result given that this population had not successfully recruited during 2014, 2015 and 2016.

Two-spined blackfish continued to be rare in the ECR, with only a few individuals being detected in the newly inundated section of the reservoir in the seven years following the commencement of filling and operational phase to date. It is likely that this species is persisting in the newly-inundated section of the reservoir, though there is no evidence to suggest that a recruiting population has yet established in the ECR. Continuation of targeted monitoring over the coming years will provide further insight into these aspects of the population of Two-spined blackfish in the ECR.

Although some other annual differences are present, the abundance and size of Rainbow trout in the ECR in 2022 was not significantly different to any other year of monitoring. Relative abundance of Brown trout was low in 2022, in contrast to the previous five years which had recorded very high abundances. Monitoring in the coming years will provide clarification if this is a true decrease in Brown trout abundance in Cotter Reservoir, or a sampling anomaly in 2022.

Small-bodied alien species other than trout continue to be detected in the ECR, with Goldfish accounting for most captures. Goldfish abundance had increased since filling commenced, most likely in response to increased availability of food resources. However, Goldfish abundance has been low since 2017, following a boom in abundance during filling and the first year of operational phase. Although Goldfish probably pose little direct threat to Macquarie perch and Two-spined blackfish, there is potential for wider effects from declines in Goldfish abundance. For instance, the loss of Goldfish within the ECR food web could see a high abundance of potential predators (cormorants and trout) needing to switch their prey consumption to Macquarie perch.

Piscivorous birds have been relatively stable in their species composition and abundance in the ECR since filling commenced, though some subtle differences in distribution have occurred. There has been an increased number of Great cormorants and Little pied cormorants in sections (primarily

section 4) that contain nesting sites and associated roosts. Breeding colonies of cormorants have far higher energy requirements than non-breeding colonies and the establishment of a breeding colony of cormorants in the ECR could increase predation pressure on adult and juvenile Macquarie perch. Cormorant management activities were undertaken as part of the Cormorant management strategy in 2014 and 2015, with mixed results. Cormorant thresholds have been revised (raised) to better reflect the increase in shoreline of the ECR.

For the first time since filling commenced, emergent macrophyte stands were detected along the shoreline of Cotter Reservoir. Three small stands of Cumbungi (*Typha domingensis*) were found, two on the eastern shoreline near the old boat ramp road, and one on the western shoreline near Bracks Hole. The stable reservoir levels over the past two years have provided conditions suitable for establishment of these emergent macrophyte stands. Whilst these stands, due to their relatively small size, are not likely to offer significant cover for adult Macquarie perch, it is heartening that establishment of emergent macrophytes is occurring in the enlarged reservoir.

Food resources of Macquarie perch (primarily decapods and microcrustaceans) showed small differences between baseline, filling and operational phases. Decapods were in low abundance in spring in both baseline and filling phases. However, there was no discernible difference in autumn decapod abundance between baseline and filling phase. There was, however, a sharp decrease in decapod abundance in autumn during the early operational phase monitoring, which was of concern as this is an important dietary item of Macquarie perch in the ECR. Monitoring since 2018 suggests that decapod abundances are returning to that observed in the baseline phase. Microcrustaceans demonstrate varying patterns through season and phase, though were in very low abundances in the latest samples. Operational phase monitoring has detected a downward trend in relative abundance of Cladocera, which have been shown to be part of Macquarie perch diet. The mechanism underpinning the reduction in Cladocera relative abundance may be related to a reduction in available resources (food and habitat) compared to baseline and filling phase.

RECOMMENDATIONS

The failure to catch Two-spined blackfish in the Bendora Reservoir reference site is the first time in 30 years of sampling this reservoir with fyke nets in the previous monitoring year (2021), and the continuing low abundances in recent years warrants further investigation. If the site continues to return very low abundances of blackfish another reference site will need to be sampled (Corin Reservoir).

Based on the detection of emergent macrophytes in autumn 2022, it is recommended that formal surveys of emergent macrophytes commence. We propose that monitoring of emergent macrophytes occur annually, during later summer early autumn.

No other change to the current monitoring program or management actions are recommended at this stage. Continued close scrutiny of adult Macquarie perch size and abundance and the annual occurrence of recruitment to YOY is recommended alongside monitoring of pest fish species such as trout and Goldfish.

BACKGROUND

Ongoing drought and its threat to water security in the ACT resulted in the recommissioning and augmentation of Cotter Reservoir from ~4 GL to 76.2 GL capacity. The enlarged Cotter Reservoir (ECR) and Cotter River upstream to Bendora dam contain four threatened fish and crayfish species: Macquarie perch *Macquaria australasica*, Trout cod *Maccullochella macquariensis*, Two-spined blackfish *Gadopsis bispinosa* and Murray River crayfish *Euastacus armatus*. Trout cod are not present in the ECR, with Murray River crayfish only confirmed from a handful of occasions. Both species are rarely encountered in the Cotter River below Bendora dam. Consequently, the major focus for threatened fish research and mitigation projects associated with the ECR has been Macquarie perch and Two-spined blackfish. Potential impacts of the construction of the ECR have been well described and reviewed (Lintermans 2005a, ACTEW Corporation 2009a, b, Lintermans 2012) and in response to these impacts, a range of projects including a fish monitoring program commenced (ACTEW Corporation 2009b).

The broad scope of the potential impacts of the ECR are summarised below.

The main threats to the Macquarie perch population in the Cotter Reservoir as a result of the ECR are related to:

- loss of adult shelter habitat (fringing emergent reedbeds)
- alteration to primary food resources associated with fringing reedbeds,
- increased predation from cormorants and trout,
- loss of riverine spawning habitat through inundation of existing habitat and restricted access to alternative habitat,
- impacts associated with competition, predation and disease transmission from existing alien fish species, and
- invasion by two additional alien fish species (Redfin perch *Perca fluviatilis* and Carp *Cyprinus carpio*).

The anticipated trophic upsurge that occurred in the newly filled ECR was considered to likely result in enhanced populations of trout within the ECR, whose impacts then spill over into the river as trout move into the river to spawn (Lintermans 2012, Todd *et al.* 2017). Threats to the riverine Macquarie perch and Two-spined blackfish populations between the ECR and Bendora dam are:

- increased predation from trout
- loss of riverine spawning habitat through inundation of existing habitat and restricted access to alternative habitat, and
- invasion upstream by two additional alien fish species (Redfin perch and Carp) should these species establish in the reservoir (Lintermans 2012).

As well as enhancing trout populations, the trophic upsurge in the ECR was considered likely to benefit Macquarie perch in the reservoir, both in terms of individual fish condition and population size, potentially providing a window of opportunity for the establishment of additional populations of this species outside the lower Cotter catchment (Lintermans 2013, Todd and Lintermans 2015). As the reservoir has filled and has been in the operational phase since 2016, the window of trophic upsurge is now considered to have largely closed.

Consequently, information on the condition and size of alien and native fish populations (including a range of life history phases), cormorants, and habitat conditions in the ECR and the river upstream is an essential requirement to adaptively manage the aquatic resources of the lower Cotter catchment.

The baseline phase of the ECR monitoring program (2010-2013) has been completed (Lintermans *et al.* 2013), which along with other available datasets provides a pre-filling baseline of threatened and alien fish species abundance and occurrence both in the impoundment and the river upstream. The filling phase of the monitoring program was conducted between 2013 – 2015 and the operational phase monitoring commenced in 2016. The underlying sampling design and priority knowledge gaps for the filling and operational phases of the monitoring program were revised and modified and now address ten management questions:

1. Has there been a significant change in the abundance and body condition of Macquarie perch in the enlarged Cotter Reservoir (Young-of-Year, juveniles and adults) as a result of filling and operation?
2. Has there been a significant change in the abundance, body condition and distribution of the Macquarie perch in the Cotter River above and below Vanitys Crossing as a result of the filling and operation of the ECR?
3. Have Two-spined blackfish established a reproducing population in the enlarged Cotter Reservoir and are they persisting in the newly inundated section of the Cotter River?
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5. Has there been a significant change in the abundance and size composition of trout in the Cotter River upstream of the enlarged Cotter Reservoir as a result of the filling and operation of ECR?
6. Are Two-spined blackfish and Macquarie perch present in trout stomachs in the Cotter River?
7. Has there been a significant change in the abundance and distribution of non-native fish species) in the enlarged Cotter Reservoir as a result of filling and operation?
8. Has there been a significant change in the abundance, distribution and species composition of piscivorous birds in the vicinity of the enlarged Cotter Reservoir as a result of filling and operation?
9. Have macrophyte beds re-established in the enlarged Cotter Reservoir?
10. Are there adequate food resources (particularly decapods) for the Macquarie perch following the filling and operation of the enlarged Cotter Reservoir?

The integrated monitoring program has field activities often addressing multiple questions (Table 1).

Table 1. Monitoring questions to be addressed at each monitoring site (see Figure 1 for location of monitoring sites).

Site	Question addressed
Cotter Reservoir	1, 3, 4, 7–10
Bracks Hole*	2, 3, 5, 6
Downstream of Vanitys Crossing	2, 5, 6
Vanitys Crossing	2, 5, 6
Spur Hole	2, 5, 6
Pipeline Rd. Crossing	2, 5, 6
Burkes Ck. Crossing	2, 5, 6
Bendora Reservoir**	3, 4
Kissops Flat***	1, 2
Cotter Hut	5, 6

*Bracks Hole has been inundated. This site has been replaced by the Downstream of Vanitys Crossing site for questions based on riverine habitats (Questions 2, 5 and 6).

** Reference site for Questions 3 and 4.

***Reference site on the Murrumbidgee River for Questions 1 and 2.

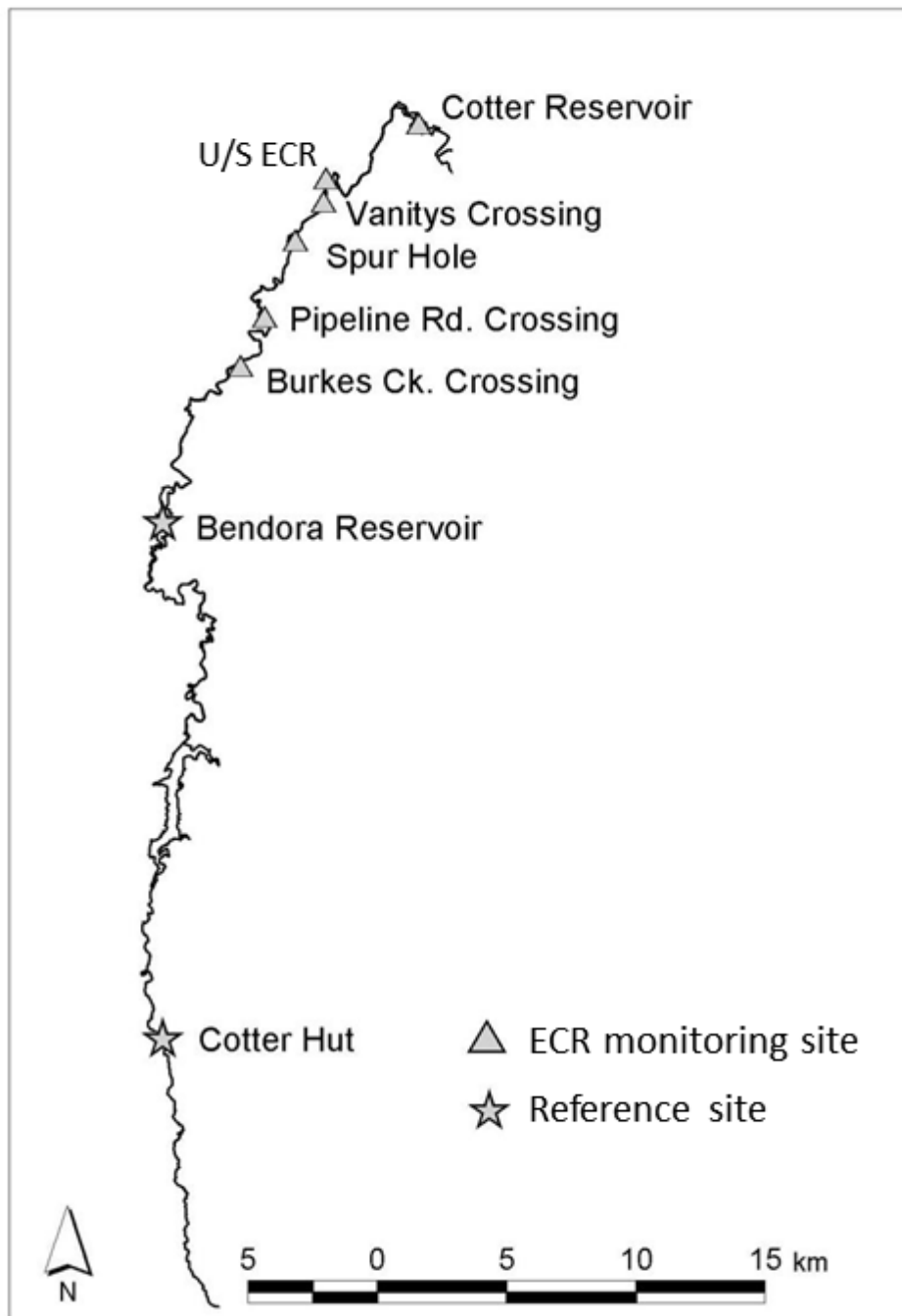


Figure 1. Location of sampling sites on the Cotter River. Note: Map does not include the reference site on the Murrumbidgee River (Kissops Flat).

The filling and operational monitoring program effectively utilised the methods and sites from the baseline monitoring program with the following changes:

- Some reference sites (Lake Ginninderra, Micalong Creek and Corin Reservoir) were excluded as a cost-saving measure, as requested by Icon Water.
- The data collected for Questions 1 and 2 has been expanded to include weight, body depth and body width of adult Macquarie perch captured (for body condition estimates).
- Snorkelling of the river immediately upstream of the ECR to Vanitys Crossing has been added to determine the recruitment input from the reservoir population of the ECR (as opposed to potential supplementation from riverine reaches further upstream)
- Boat electrofishing is being trialled to determine its effectiveness in capturing adult Macquarie perch compared to gill netting
- The site immediately downstream of Bendora Dam has been removed from the program due to budget restrictions.
- For questions 2, 5 and 6, the site at Bracks Hole (immediately upstream of the old Cotter Reservoir full supply level) has been replaced by another riverine site immediately upstream of the full supply level of the ECR (but downstream of Vanitys Crossing). This replacement was necessary as Bracks Hole has already been inundated and no longer represents a riverine site. A riverine monitoring site immediately upstream of the full supply level of the ECR is required as this is the most likely area where impacts of the operation of the ECR will be greatest.
- Bait traps have been added to the sampling techniques for Question 3 and Question 7. The addition of bait traps will increase the likelihood of capture of juvenile Two-spined blackfish (detection of recruitment) and also increase the likelihood of capture of small and juvenile non-native species associated with Question 7.
- The intensity of sampling effort for characterisation of trout diet associated with Question 6 has changed, as has the way in which the stomach contents of trout are processed (now field coarse visual inspection rather than laboratory dissection), as requested by Icon Water.
- The filling and operational phases monitoring program has proposed methods for assessing the establishment of macrophytes that were not covered in the baseline monitoring program.
- Additional fyke netting and gill netting effort is employed in the expanded ECR (2017 onwards).
- Additional boat electrofishing at night to be undertaken in Cotter Reservoir (2019 onwards)

The rationale and results from each of the 10 management questions are presented in the following sections.

HYDROLOGICAL SUMMARY

The Cotter River experienced prolonged drought through the late 1990-2000s (van Dijk *et al.* 2013), with the phenomenon worsening from 2006 until the latter half of 2010 when significant rains resulted in flooding (Figure 2). This flooding caused the original Cotter Reservoir to rise by 2 – 4 m from mid-October 2010 to Mid-April 2011 (Figure 3). A significant single rainfall event and associated large-scale flooding also occurred in early 2012, which led to water levels in the under-construction Enlarged Cotter Reservoir prematurely increasing by about 10 – 12 m in February 2012 (Figure 3). In terms of effects on monitoring results, 2010/11 fish monitoring reflected the previous year that was dry and the ending of an extended extreme drought. Monitoring in 2011/12 and 2012/13, and to some extent 2013/14, were years where the preceding year had an average of high rainfall and discharge, when compared to recent history. Monitoring in 2014/15 and again in 2015/2016 followed relatively dry conditions as it appeared the area was moving towards another period of lower than average rainfall. Monitoring in 2016/2017 followed a wetter than average winter and spring where all three reservoirs on the Cotter River filled in winter 2016, and remained full throughout the Macquarie perch spawning season (September – December) resulting in the Cotter River between Bendora Dam and the Enlarged Cotter Reservoir operating as largely unregulated (Figure 2). The monitoring year of 2019 / 2020 saw a return to flows which were dominated by regulated flow releases (because of dry climatic conditions), except for a few short, high flow pulses associated with rainfall events (notably early March 2020) (Figure 2). Because of these low inflows and abstraction for water supply, the Enlarged Cotter Reservoir level receded to approximately 9 m below full supply level, as of May 2020 (Figure 3). All three Cotter reservoirs filled and overtopped in approximately September 2020, and subsequent rainfall has held them full for the majority of the time since (Figure 3). This also meant that flow in the Cotter River below Bendora Reservoir was largely operating as unregulated for this period, characterised by high base flow and large peaks in discharge (notably from early November 2021 – mid January 2022), associated with large rainfall events (Figure 4). The Cotter River at Vanitys Crossing was flowing at greater than 100 ML Day⁻¹ (the maximum discharge at which representative and safe river sampling can be undertaken in this reach) for all but three days of the autumn 2022 sampling period (8- 10th April) (Figure 2).

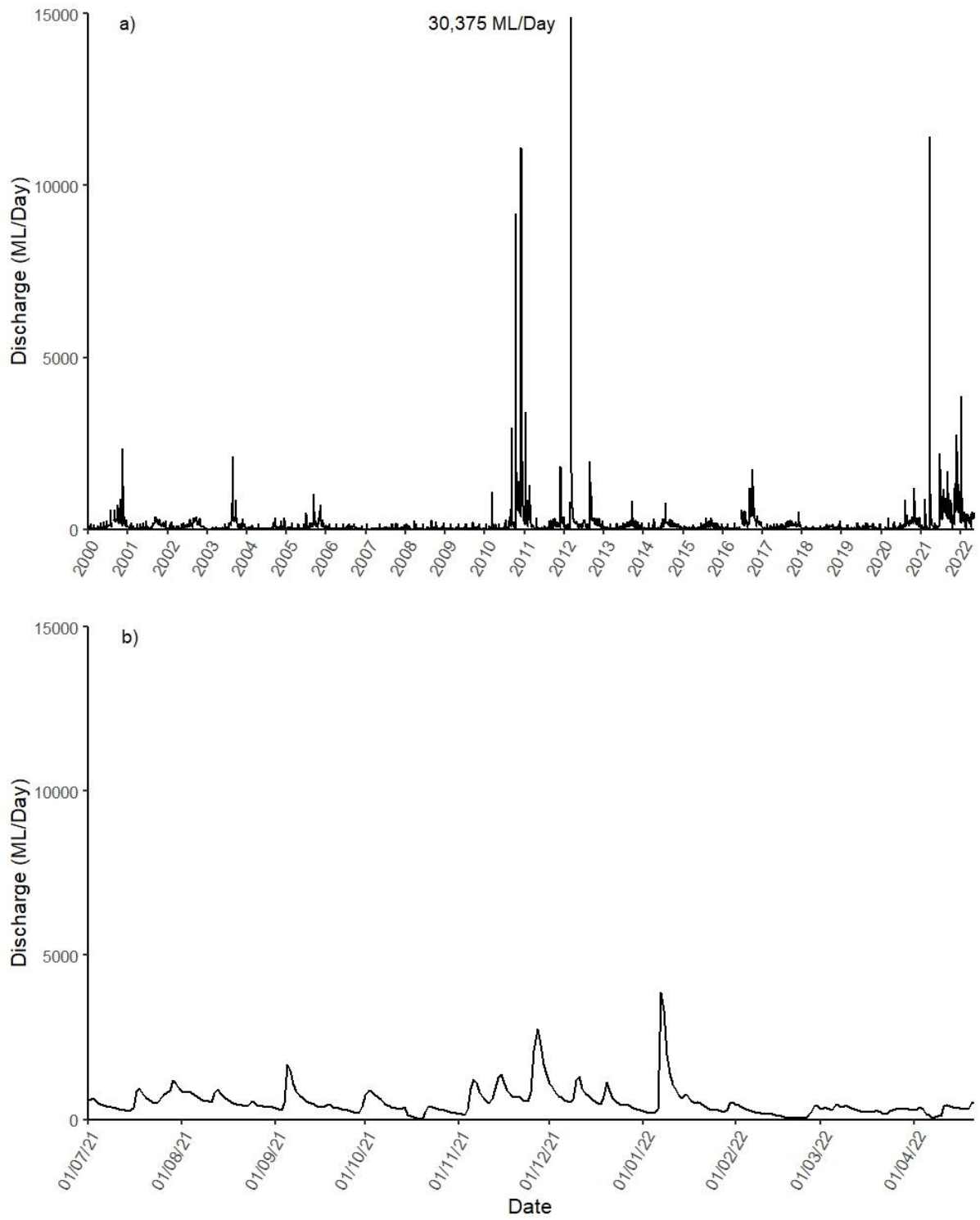


Figure 2. Daily discharge of the Cotter River at Vanity's Crossing from a) January 2000 until May 2022 and b) July 2021 – May 2022.

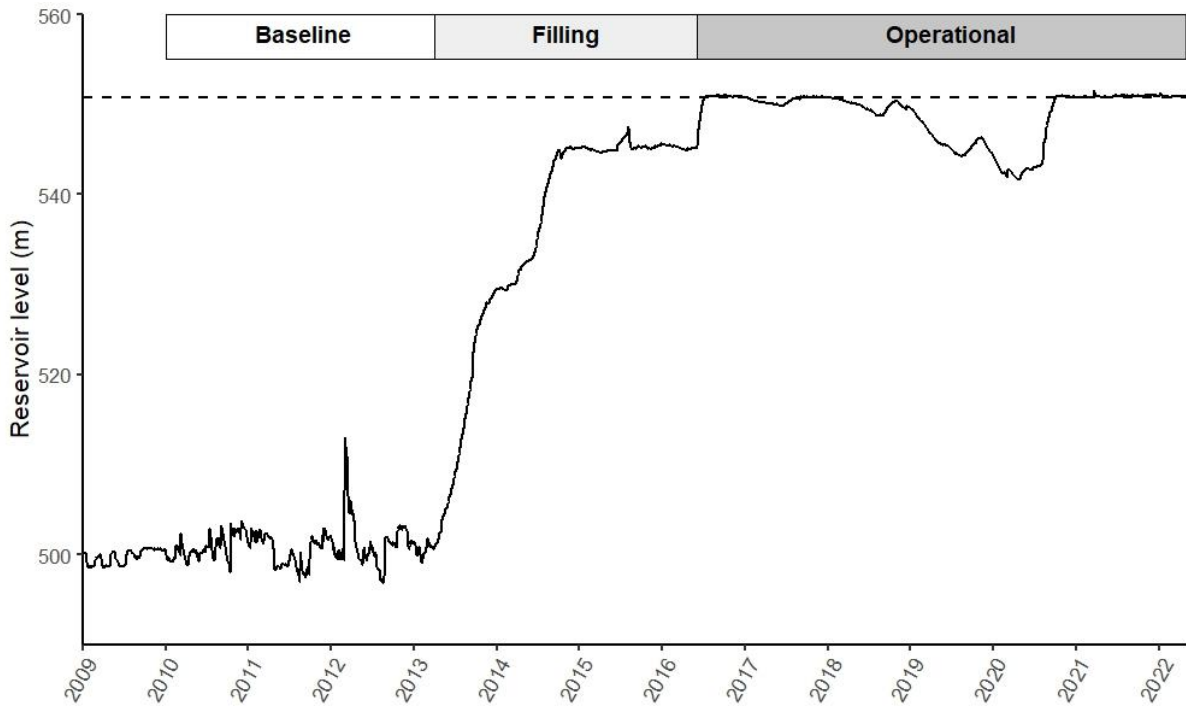


Figure 3. Water level (in metres above sea level) of Cotter Reservoir from January 2009 until May 2022. Rectangles indicate the three monitoring phases: white = Baseline, grey = Filling, hatched = Operational. Blue line indicates Enlarged Cotter Reservoir full supply level. Full supply level prior to enlargement was 500.8 m ASL.

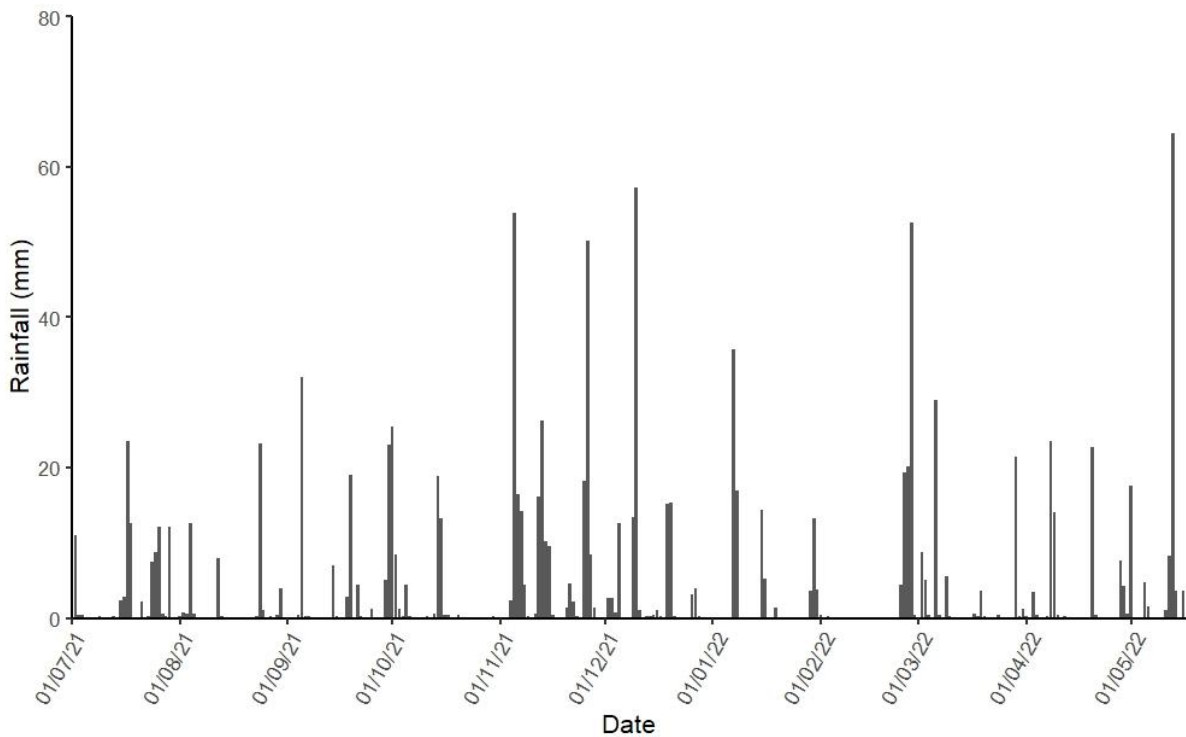


Figure 4. Daily rainfall (mm) measured at Pierces Creek meteorological station from July 2021 – May 2022.

MONITORING METHODS, RESULTS AND DISCUSSION

QUESTION 1: Has there been a significant change in the abundance and body condition of Macquarie perch in the enlarged Cotter Reservoir (young-of-year, juveniles and adults) as a result of filling and operation?

BACKGROUND

A range of potential threats such as loss of habitat, interactions with alien fish species, and predation by cormorants can impact the Macquarie perch population in the Cotter River and reservoir as a result of the filling and operation of the new ECR. In considering these potential ECR impacts, we must account for natural fluctuations in Macquarie perch abundance that can arise from interannual variations in climate, flow regime, stochastic extreme events (flood, drought, etc.) and other factors that influence rates of spawning, recruitment and mortality. Body condition of adult Macquarie perch is a key indicator of reproductive potential, and so monitoring changes in adult body condition can be a useful indicator of future recruitment events and overall population trajectories (Gray *et al.* 2000). Spawning of reservoir-resident Macquarie perch may be impacted by newly encountered riverine barriers in a filling reservoir (Broadhurst *et al.* 2016a), so early detection of spawning success (via snorkelling for larvae) and how this relates to young-of-year (YOY) captured in the reservoir via netting will all contribute to the understanding of recruitment success or failure for a given year. Previous monitoring indicated that sampling for adult Macquarie perch in a filling reservoir may be difficult using gill nets (as fringing vegetation becomes flooded) and that a complementary technique (boat electrofishing) required exploration (Lintermans *et al.* 2013).

METHODS

The sampling design for Question 1 largely follows that of the baseline monitoring program Question 1 (Table 2; (Lintermans *et al.* 2013). An additional metric – the wet weight of individuals captured in gill nets - was used to calculate fish condition. Boat electrofishing was added as a sampling method to mitigate potential sampling inefficiencies via gill netting for adult Macquarie perch during ECR filling (see below for details). To determine the likely contribution of young-of-year (YOY) recruitment to the reservoir population by reservoir adults, snorkelling of the river from immediately upstream of Cotter Reservoir (ECR) to Vanitys Crossing was undertaken.

Table 2. Outline of the sampling design for Question 1 of the ECR monitoring program.

Feature	Detail
Target species and life history phase	Macquarie perch <i>Macquaria australasica</i> . Adults (> 150 mm total length (TL)), Juveniles (100 - 150 mm TL) and young-of-year (< 100 mm TL). Larvae and early juveniles observed during snorkelling are likely to be 15 – 25 mm TL.
Sampling technique/s	Gill nets (10 (4 x 100 mm; 4 x 75 mm; 2x 125 mm stretch mesh)) per night for 5 nights, with an additional 2 x 125 mm gill nets per night to capture larger individuals) and fyke nets (12 mm stretch mesh, 20 per night for 3 nights in Cotter Reservoir; 12 per night for 2 nights at Kissops Flat). Boat-electrofishing 12 shots per shoreline per section for daytime and 6 shots per shoreline section for night-time sampling. Snorkelling (visual survey) of stream pools for larvae / early juveniles.
Timing	Netting and electrofishing was conducted annually in March - April; snorkelling in Nov/Dec.
Number / location of sites	One impacted site: Enlarged Cotter Reservoir (and river immediately upstream); one reference site: Kissops Flat (upper Murrumbidgee River). No snorkelling at Kissops Flat.
Information to be collected	Number and total length for all Macquarie perch. Wet weight (g) for subadults/adults captured in gill nets. Number of larvae/early young-of-year per pool.
Data analysis	Catch-per-unit-effort (CPUE) assessed between years using analysis of similarity (ANOSIM) for gill net data and PERMANOVA and ANOSIM for fyke net data. Length and body condition of individuals captured gill netting will be assessed between years (baseline, filling and operational) using a Kruskal Wallis ANOVA on ranks.

Non-larval sampling targeted adult, juvenile and young-of-year Macquarie perch. Individuals were classed as adults if they were > 150 mm total length (TL), based on results from Ebner and Lintermans (2007) who found that males are sexually mature from this size. At the time of netting (i.e. autumn), Young-of-year are approximately 60 – 99 mm TL based on results of the baseline data collection (Lintermans *et al.* 2013). Individuals were considered juvenile if they fell between 100 – 150 mm TL. Snorkelling surveys target larval and early young-of-year Macquarie perch that are ~ 2 – 4 weeks of age (~15 – 25 mm TL).

Sampling was conducted at two sites; ECR (impacted site) and Kissops Flat (reference sites for young-of-year and juveniles). Only fyke netting was employed at Kissops Flat (see below for details). In the ECR two sampling techniques were employed to capture a representative sample of the entire size-range of the Macquarie perch population. Both gill nets (free-floating, multi-filament) and fyke nets (12 mm stretch-mesh single-winged) were deployed, as the former is most effective for capturing

adult Macquarie perch and the latter is most effective at capturing young-of-year and juvenile Macquarie perch (Ebner and Lintermans 2007, Lintermans *et al.* 2013, Lintermans 2016).

Gill nets were deployed as per the baseline and previous filling and operational monitoring. Specifically, 12 gill nets were set independently around the perimeter of the reservoir in March 2022. The reservoir was divided into five longitudinal sections, with two gill nets set in each section. Gill netting was undertaken over five nights (based on power analysis conducted Robinson 2009), though was not conducted for more than two consecutive nights at a time to avoid stress on adult Macquarie perch by multiple sequential re-captures. In 2022, an additional two 125 mm gill nets (1 x deep-drop (66 meshes deep)) have been added to the previous effort of 10 gill nets (75, 100, 125 mm stretch mesh) to capture the increasingly larger adult Macquarie perch in Cotter Reservoir (following recommendations in Broadhurst *et al.* 2018). Gill nets were set for six hours soak time commencing at ~15:30hrs following the existing threatened species netting protocol to minimise potential issues with prolonged retention of threatened fish in gill nets.

Twenty fyke nets were set singularly around the perimeter of the reservoir over three nights in March 2022. Twelve fyke nets were set for two nights in the pool at Kissops Flat in February 2022. Fyke nets were set for ~16-hour soak time (existing fyke netting protocol) commencing at ~15:30-16:00 hrs.

Larval monitoring was not undertaken in 2021, as river levels were far too high to safely undertake snorkelling surveys. Cotter River discharge (recorded at Vanitys Crossing) varied between ~600 – 2700 ML Day⁻¹ from mid-November to mid-December 2021.

The baseline monitoring report suggested that an alternative sampling technique for adult Macquarie perch was required during the filling phase as gill nets failed to capture any adult Macquarie perch during high water levels in a flooded reservoir in 2012, most likely due to gill nets being set on partially-submerged vegetation further from the shoreline than usual (Lintermans *et al.* 2013). Boat electrofishing occurred across multiple days with the reservoir divided into five longitudinal sections, with twelve 90-second “on time” electrofishing shots undertaken along each shoreline (left and right banks) of each section (10 replicates in total). Catches from boat electrofishing are compared with gill netting results from the same year to determine if catches of adult Macquarie perch follow the same patterns between techniques. To attempt to increase captures of large adult Macquarie perch (as per recommendations from Broadhurst *et al.* 2018), a trial of night-time boat electrofishing commenced in 2019. Night-time electrofishing mimicked daytime, although at a reduced effort per shoreline section (six shots per sections instead of 12). As this was an increased survey aimed at capturing adult Macquarie perch, individuals less than 150 mm TL were observed only (not counted), although broad estimates of abundance were made for individuals < 100 mm TL (young-of-year) and Juveniles (100 > 150 mm TL) for each shot. Because of high water turbidity in 2021 (associated with high rainfall, runoff and river inflows), no boat electrofishing was undertaken.

Abundance of Macquarie perch was standardised for effort applied during each sampling technique by calculating number of fish caught per hour (i.e., catch-per-unit-effort, or CPUE). Given the spatial ecology of Macquarie perch, CPUE of gill netted Macquarie perch was then scaled according to the shoreline length at the time of sampling, which varies with ECR water level. This was done by multiplying the CPUE for each net night by the proportional increase in shoreline according to the

reservoir water level in each survey year (relative to the old Cotter Reservoir water level above sea level). In the case of fyke netting, where net effort was also increased from 2017 onwards, the increased shoreline was divided by the increased proportional net effort for these years. See below for scaled CPUE equation:

$$\text{Scaled CPUE} = \text{CPUE} / (\text{Prefilling ECR shoreline} / \text{shoreline at time of sampling}) / (\text{baseline number of nets} / \text{current number of nets}).$$

Analysis of Macquarie perch CPUE in gill nets (excluding the additional 125 mm gill nets used in 2022 from analyses) was assessed between years using analysis of similarity (ANOSIM) with phase as a fixed factor and year as a random factor nested within phase. Gill netting data was $\text{Log}_{10}(x+1)$ transformed and fyke netting data was $\text{Log}_{10}(x+1)$ transformed to deal with skew, and then a resemblance matrix was constructed using the modified Gower (base 2) dissimilarity measure for gill netting data and a modified Gower base 2 (+ dummy variable to deal with double-zeros across sample pairs) for fyke netting data. Tests were run with a maximum of 9999 permutations. For fyke net data, size classes (<100 TL, >100mm TL) were included as variables, with site and phase as fixed factors, and a random factor of year nested within phase for a maximum of 9999 permutations. To test between differences in Macquarie perch CPUE in fyke nets for each size class (<100 mm TL and >100 mm TL), PERMANOVA using Type III sum of squares in a repeated measures design was employed and used for pairwise tests (site and year as fixed factors) (following Anderson et al. (2008)). This approach allowed for an unbalanced design arising from the different number of samples collected across years. Significant interactions were interpreted using threshold metric MDS performed on group centroids for site by year. Graphical presentations of site-level means with 95% confidence limits (with Bonferroni corrections applied for $n = x$ sampling years) were then used to the magnitude of pairwise variations in CPUE of Macquarie perch size classes among sites and years. Condition of adult Macquarie perch was analysed using Fulton's condition index, which is calculated as $K = 100(\text{weight}/\text{length}^3)$ following (Ricker 1975). Size (TL) and body condition of the adult population was analysed using Kruskal-Wallis ANOVA tests to determine if a significant change occurred through time. Pairwise comparisons were then undertaken using Dunn's method. ECR monitoring program body condition data was compared against historical data from 2007 – 2009 (data from Lintermans *et al.* 2010).

RESULTS

Adult Macquarie Perch

A total of 19 Macquarie perch were captured using all gill nets in ECR in 2022, which ranged from 94 – 405 mm TL (Figure 5). Adult Macquarie perch have been captured by gill nets in every year since monitoring began in 2010. Adult Macquarie perch abundance was highest in 2015, more than double the next most abundant years (2014, 2016 and 2017, and roughly quadruple the abundances of all other years, including 2022 (Figure 6). Macquarie perch CPUE was significantly different among years (Global $R = 0.013$, $p < 0.01$), with 2015, 2016 and 2017 having significantly higher captures of adult Macquarie perch compared to all other years. There was no significant difference in CPUE among monitoring phases (Global $R = 0.126$, $p = 0.227$).

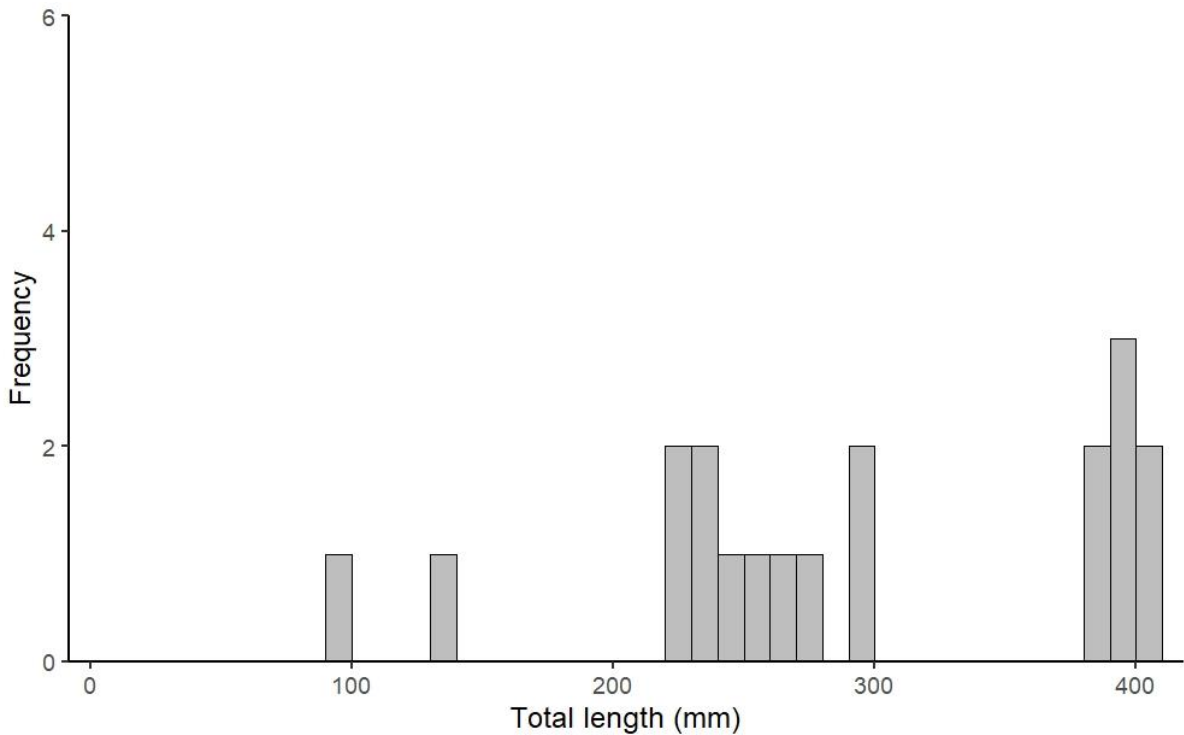


Figure 5. Length frequency of Macquarie perch captured from the Enlarged Cotter Reservoir in autumn 2022 using gill nets.

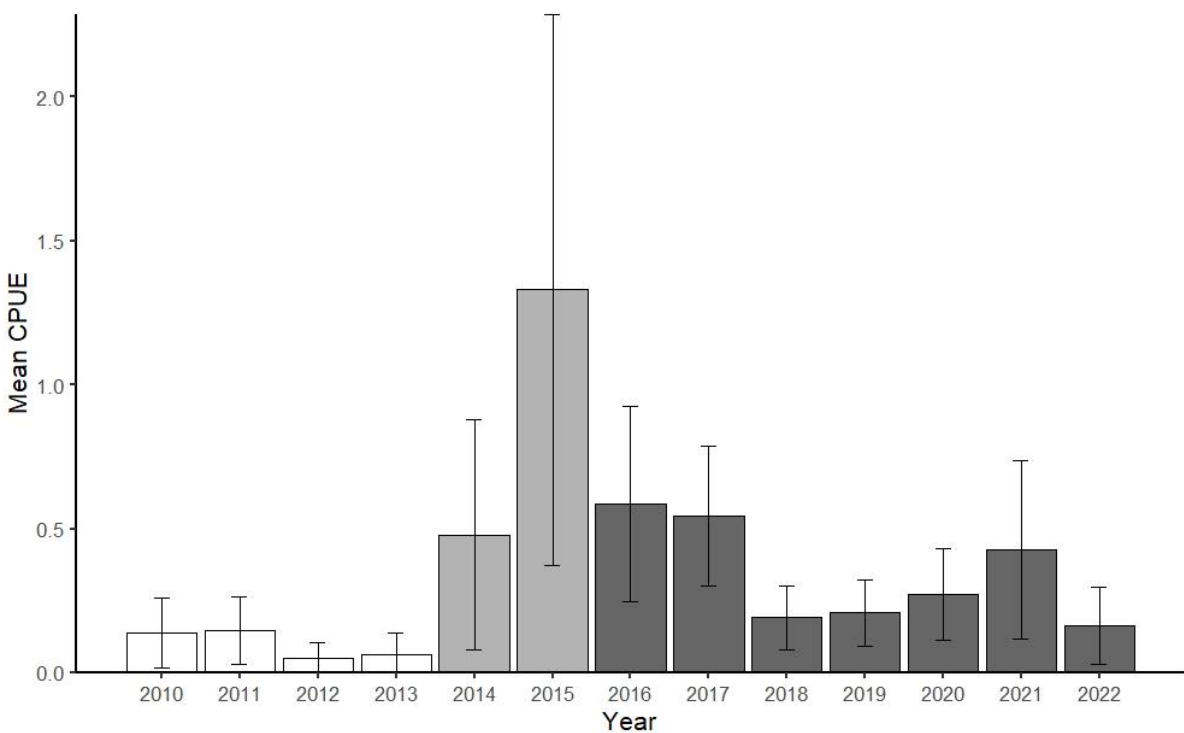


Figure 6. Relative abundance (displayed as mean CPUE \pm 95% Confidence limits with Bonferroni correction, and scaled to relative reservoir shoreline length at time of sampling) of adult Macquarie perch captured in Cotter Reservoir using gill nets between 2010 – 2022. White bars indicate baseline phase, light grey bars indicate filling phase and dark grey bars indicates the operational phase of the Enlarged Cotter Reservoir (ECR); bars are arranged in chronological order from 2010 to 2022 from left to right on the x-axis.

Length of adult Macquarie perch captured in gill nets was significantly different between years. For the most part, individuals captured in baseline and filling phases were significantly smaller than those captured in operational phase ($H_2 = 75.809$, $P < 0.01$) (Figure 7). Length of Macquarie perch captured in gill nets in 2022 was not different to any of the baseline years monitored (

Table 3, Figure 5 and Figure 7). Length of adult Macquarie perch captured in gill nets in 2022 was largely bimodal, with one mode between 220 – 300 mm TL and the other mode between 380 – 410 mm TL (Figure 5). Condition of adult Macquarie perch captured in gill nets was significantly higher during the filling phase (2014 – 2015) and operational phase (2016– 2022) compared to baseline (2007 – 2009) phase ($H_2 = 51.271$, $P < 0.01$). Mean Fulton’s condition index declined between 2017 and 2020, though has stabilised in the years since (Figure 8).

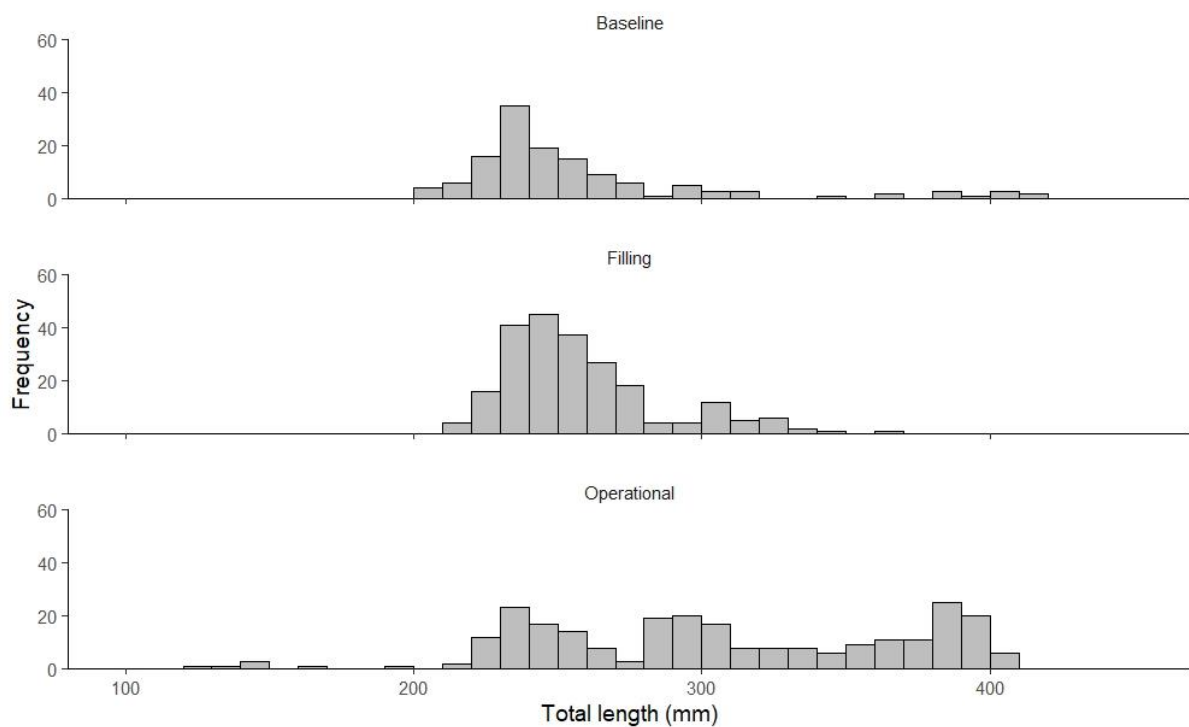


Figure 7. Length frequency of all Macquarie perch captured in gill nets in Cotter Reservoir for each monitoring phase from 2010 to 2022.

Table 3. Raw numbers of Macquarie perch in each size class captured in Cotter Reservoir using gill nets each year over the period 2010 – 2022. Monitoring phases are indicated by shading: none = Baseline; blue = Filling; green = operational.

Size classes (TL)	2010	2011	2012	2013	2014	2015	2016	2017	2018	2019	2020	2021	2022
< 300 mm	39	41	13	18	66	120	23	10	8	5	12	33	12
300 > 349 mm	2	6	1	2	5	26	33	21	1	2	1	0	0
350 >369 mm	1	0	0	0	0	1	1	11	0	0	2	0	0
>370 mm	4	3	4	0	0	1	2	14	10	14	15	9	7
All sizes total	46	50	18	20	71	149	59	56	20	21	33	42	19
<i>% of 200-300mm</i>	85	82	72	90	93	81	39	18	40	24	36	79	52

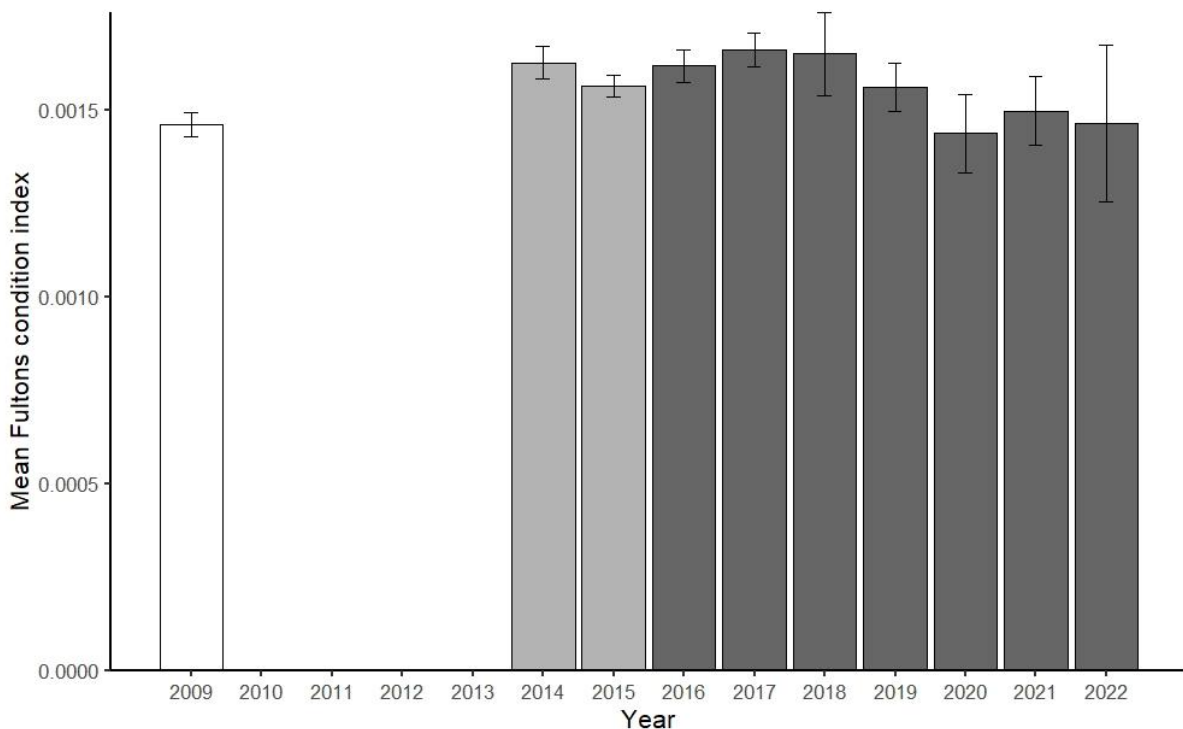


Figure 8. Mean annual condition (using Fulton’s condition index) of adult Macquarie perch captured in gill nets in Cotter Reservoir for from 2009 to 2022. White bars indicate baseline phase, light grey bars indicate filling phase and dark grey bars indicates the operational phase of the Enlarged Cotter Reservoir (ECR); bars are arranged in chronological order from 2009 to 2022 from left to right on the x-axis.

Boat electrofishing

A total of forty-four adult Macquarie perch were captured by boat electrofishing in 2022 ranging in lengths from 201 – 425 mm TL (Figure 11). As for the gill netting captures, length frequency of adults caught by electrofishing was bimodal, with modes around 250 mm and 400 mm TL (Figure 11). Of these, 23 individuals were captured during the day and 21 individuals captured at night. Relative abundance of adult Macquarie perch captured by boat electrofishing in 2022 was among the highest since this method was employed in 2014 (Figure 12).

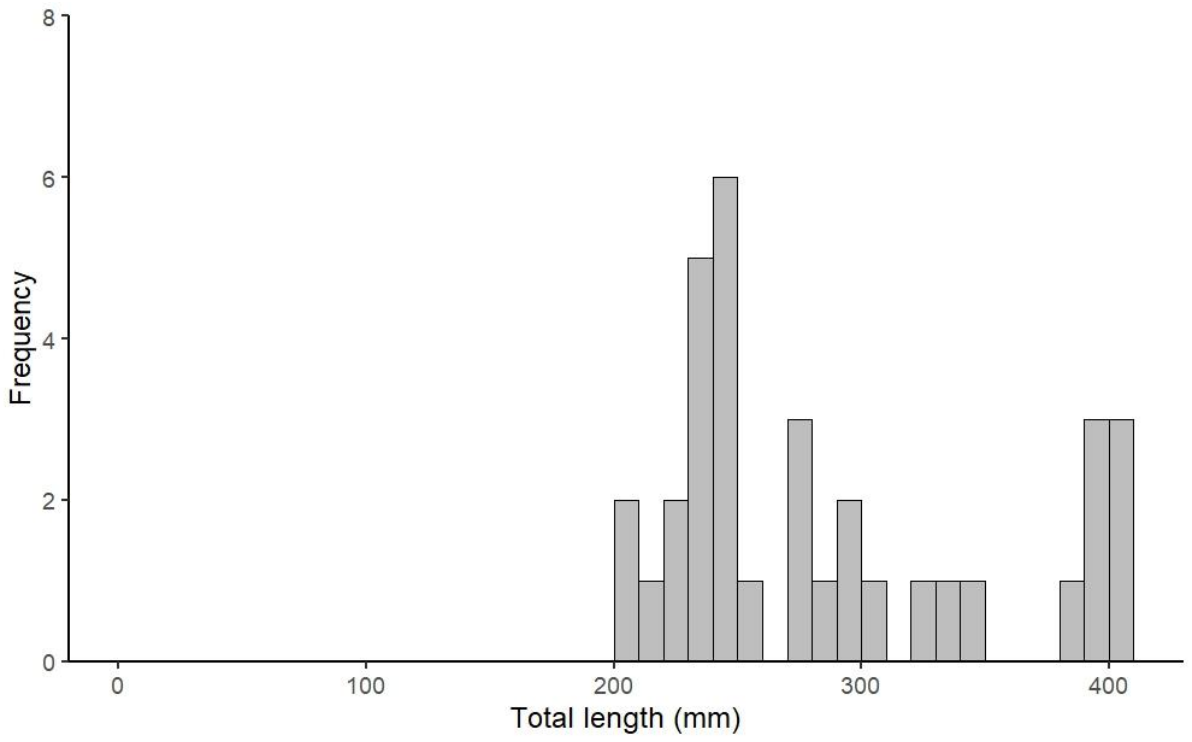


Figure 9. Length frequency of all Macquarie perch captured via boat electrofishing in Cotter Reservoir in 2022.

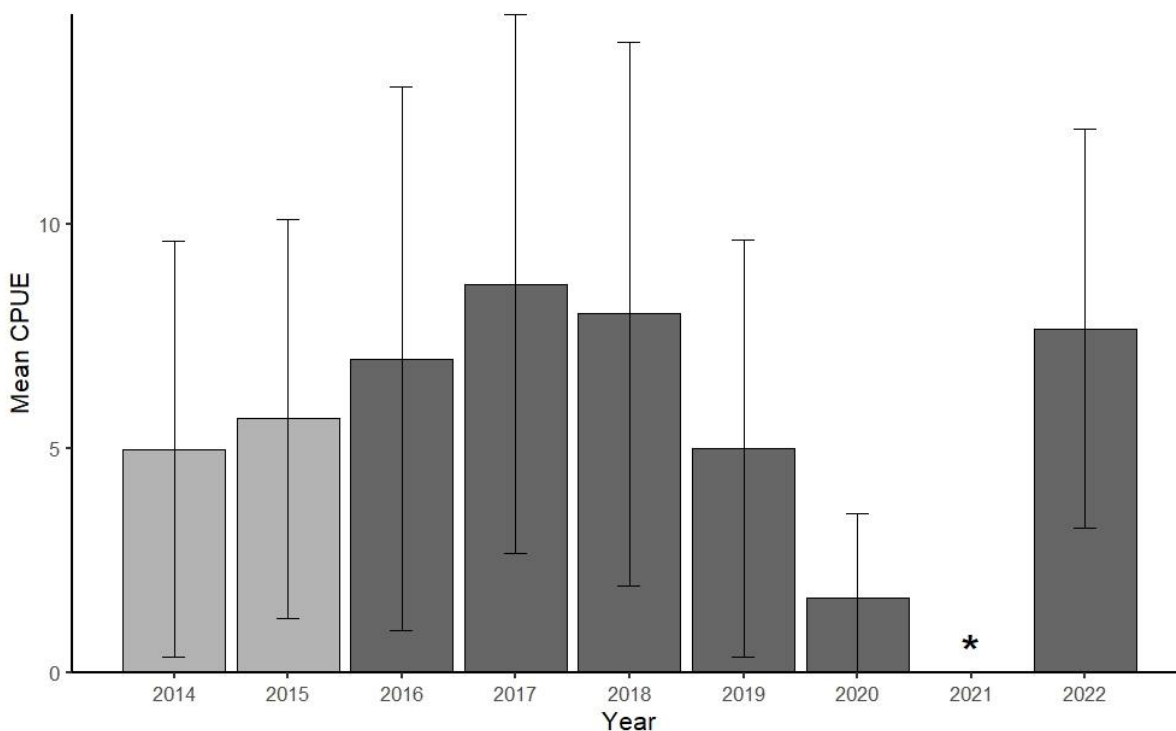


Figure 10. Relative abundance (displayed as mean CPUE \pm 95% Confidence limits with Bonferroni correction) of adult Macquarie perch captured in Cotter Reservoir via boat electrofishing between 2014 – 2022. Light grey bars indicate filling phase and dark grey bars indicates the operational phase of the Enlarged Cotter Reservoir (ECR); bars are arranged in chronological order from 2014 to 2022 from left to right on the x-axis. * Boat electrofishing not undertaken in 2021 due to high water turbidity.

Fyke netting - all age classes

A total of 364 Macquarie perch ranging from 40 – 285 mm TL were captured in 2022 from the ECR using fyke nets (Figure 11). CPUE of Macquarie perch (all sizes combined) captured by fyke netting were significantly different between the sites, years and phases, with a significant site by phase interaction (Table 4). The significant site by year interaction is largely driven by the lack of young-of-year at Cotter Reservoir during 2014 – 2016 (Figure 12).

Table 4. Results of PERMANOVA comparison of catch-per-unit-effort of Macquarie perch (all sizes combined) in fyke nets deployed in Cotter Reservoir and Kissops Flat each year over 2010 to 2022 (bold text indicates significant effects at the P(permanova) 0.05 level).

Source	df	SS	MS	Pseudo-F	P(permanova)	Unique perms
Site	1	0.66875	0.66875	23.008	0.0001	9944
Phase	2	0.78867	0.39434	6.3594	0.008	9662
Year(Phase)	10	0.70061	0.070061	2.4105	0.0009	9907
Site x Phase	2	0.16041	0.080204	2.7594	0.0311	9956
Residuals	844	24.531	0.029065			
Total	859	26.888				

Juvenile Macquarie perch

In 2022 a total of 271 Macquarie perch juveniles / sub adults (> 100 mm TL) were captured in fyke nets. Juvenile Macquarie perch have been captured each year since monitoring began in 2010 but were particularly low in abundance during 2015 – 2017 as a result of successive years of recruitment failure over 2014 – 2016. There was no significant difference in the relative abundance of juvenile Macquarie perch between sites (Global R = -0.024, p = 0.999), or years (Global R = 0.007, p = 0.071) (Figure 12).

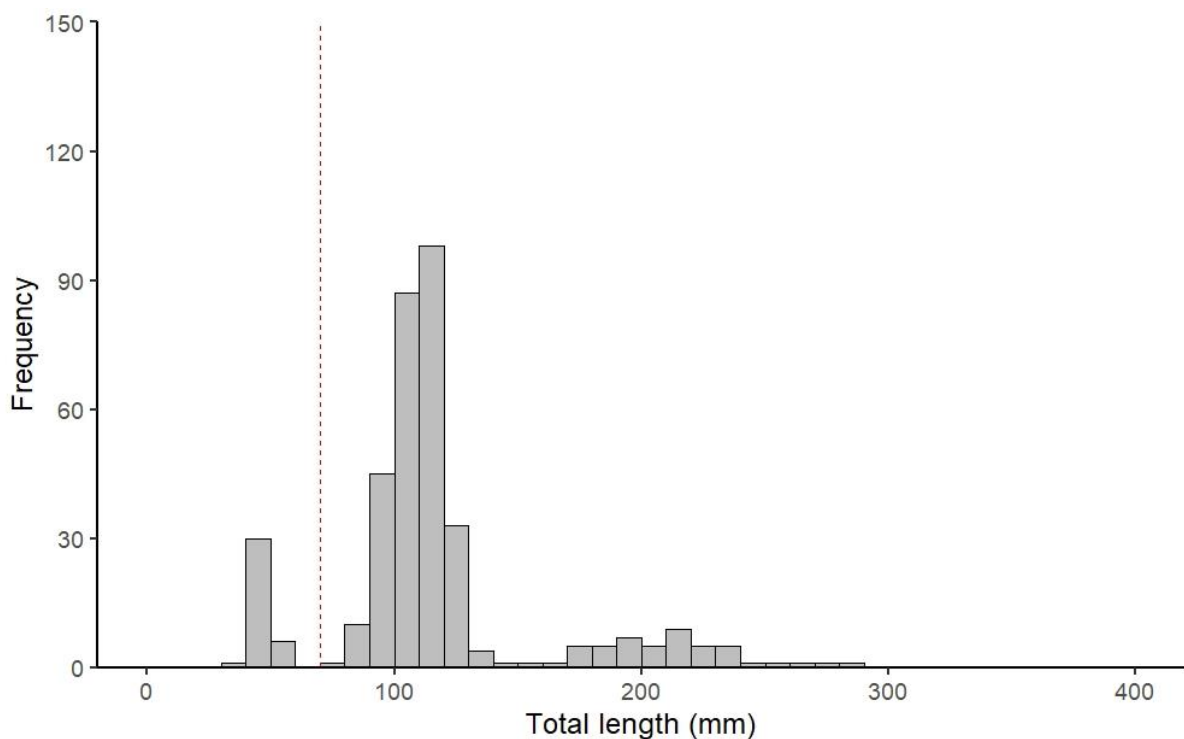


Figure 11. Length frequency of Macquarie perch captured from the ECR in autumn 2022 using fyke nets (red dashed line indicates cut-off for length of young-of-year individuals for the monitoring year).

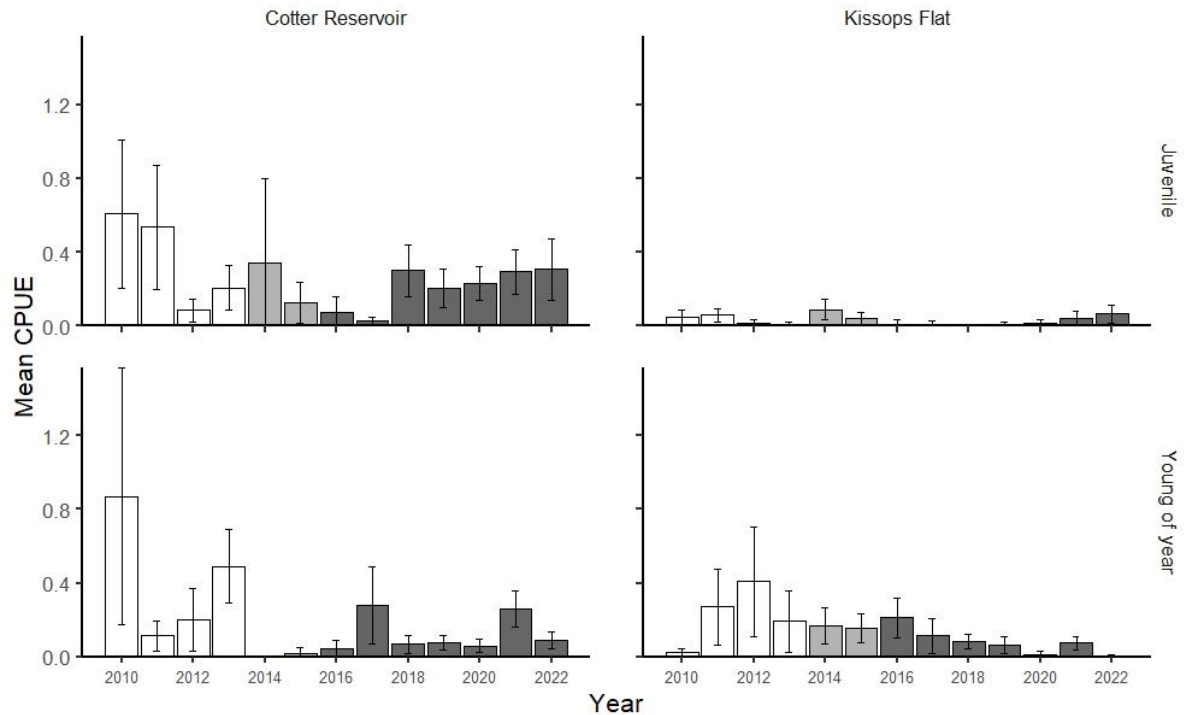


Figure 12. Relative abundance (displayed as mean CPUE \pm 95% confidence limits with Bonferroni corrections, scaled to relative net effort versus shoreline length at the time of sampling) of juvenile (>100 mm TL) and young-of-year (< 100 mm TL) Macquarie perch captured in Cotter Reservoir (pre and post enlargement) (impact site) and Kissops Flat (reference site) using fyke nets between 2010 and 2022. White bars indicate baseline phase, light grey bars indicate filling phase and dark grey bars indicates the operational phase of the Enlarged Cotter Reservoir (ECR); bars are arranged in chronological order from 2010 to 2022 from left to right on the x-axis. *Young-of-year Macquarie perch*

A total of 37 young-of-year (YOY) Macquarie perch were captured using fyke nets in the ECR in 2022. Operational years 2017 – 2022 show an improvement in CPUE of YOY compared to filling 2014 – 2015 and early operational year 2016 when extremely low abundances of YOY were captured (Figure 12). There was no significant difference in the CPUE of YOY Macquarie perch between sites (Global R = -0.01, $p = 0.880$), but there was a significant difference between years (Global R = 0.031, $p < 0.01$). Pairwise comparisons among years suggests a mixture of differences, including those between pairs of recent pre- and post-filling years. Captures of YOY Macquarie perch in the reservoir in 2022 were the same as all other years except 2010, which had significantly high abundance compared to most other years (Figure 12). YOY Macquarie perch were detected in all years of monitoring at the reference site (Kissops Flat, upper Murrumbidgee River) using fyke nets but were the lowest on record at this site since monitoring began with only two individuals captured (Figure 12 and Figure 13).

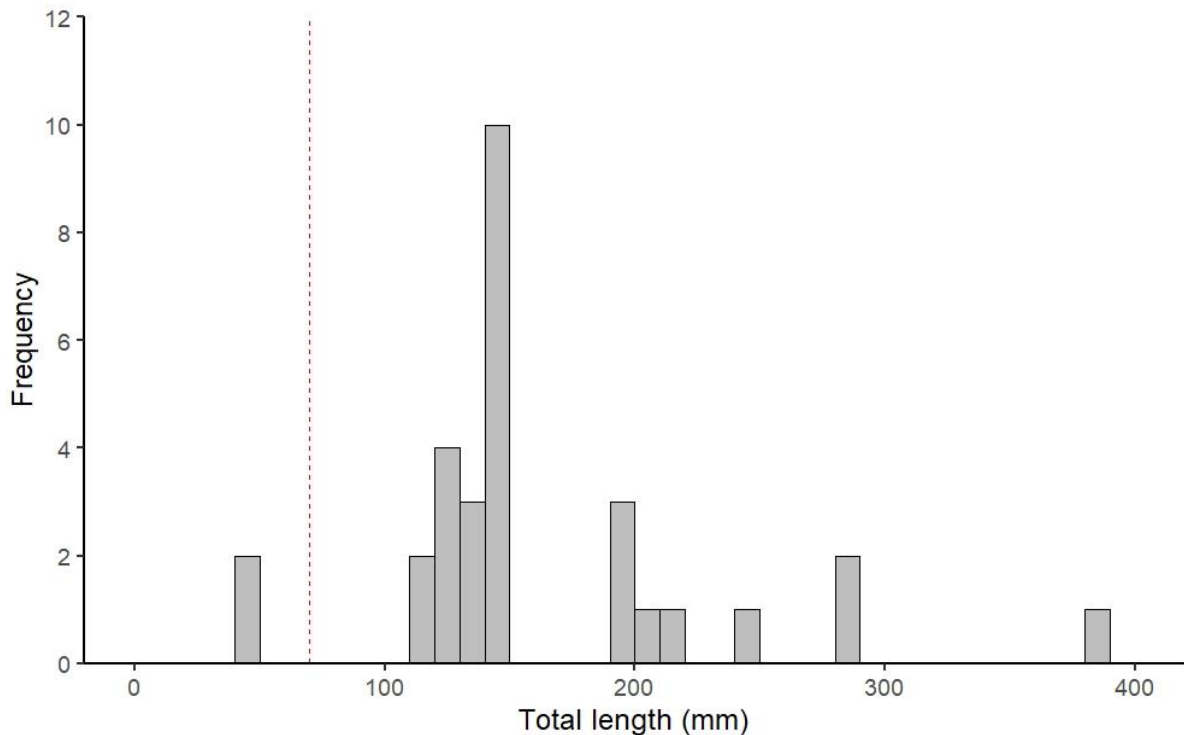


Figure 13. Length frequency of Macquarie perch captured from Kissops Flat on the upper Murrumbidgee River in autumn 2022 using fyke nets (red dashed line indicates cut-off for length of young-of-year individuals).

DISCUSSION AND CONCLUSIONS

Annual abundances of Macquarie perch size classes in Cotter Reservoir are highly variable since monitoring began, which is partly expected due to natural variations in recruitment and mortality arising from a range of environmental factors. However, the question is to what extent more recent changes in the size- structure of fish can be attributed to shifts in the reservoir habitat conditions, and connectivity to upstream sections of the Cotter River. Most recently, concerns were raised about the failure of Macquarie perch recruitment over the 2014 – 2016 period since filling of the ECR began, as indicated by very low or nil catches of YOY Macquarie perch in the ECR over that period, and very low juvenile abundance in 2017. More recent monitoring from 2017 – 2022 has indicated successful spawning and recruitment to YOY, with captures of this size class higher than some of the baseline monitoring years prior to the commencement of filling. Notably, there is a strong class of 1+ year old (juvenile) fish captured in the last four years (2018 – 2022), suggesting good annual recruitment conditions through to 1 – 3-year old individuals.

Relative abundance of adult Macquarie perch captured in 2022 in gill nets was the lowest since 2013. In isolation, this may be some cause of concern, though when coupled with the other survey methods used in Cotter Reservoir, the number of adult Macquarie perch (> 200 mm TL) captured in 2022 was among the highest since 2014 (Table 5). The relatively high abundance of adults present in the reservoir, though low capture of adults in the gill nets in 2022 indicates a change in the

catchability of adult Macquarie perch in 2022. A possible explanation is that adult Macquarie perch movement behaviour or habitat use may have changed (moving less to feed, occupying deeper water) in autumn 2022 (during abnormally high autumn river flows), rendering adults less likely to encounter the gill net fleet.

Table 5. Raw number of Macquarie perch > 200 mm TL captured using gill nets, fyke nets and boat electrofishing in Enlarged Cotter Reservoir 2014 – 2022. *Boat electrofishing was not undertaken in 2021 because of high water turbidity.

Year	Boat e-fish	Fyke	Gill	Total
2014	2	4	71	77
2015	16	24	152	192
2016	21	3	59	83
2017	15	5	59	79
2018	12	4	19	35
2019	7	3	21	31
2020	5	21	30	56
2021	*	47	42	89
2022	44	29	17	90

Adult Macquarie perch captured in both filling and operational phases displayed significantly higher body condition relative to individuals captured in baseline phase. Condition of adult Macquarie perch has declined since early filling phase years, though seems to have stabilised since 2020. Body condition was not measured during baseline monitoring from 2010 – 2013. Higher body condition was expected as rising water levels of Cotter Reservoir inundated banks and vegetation, resulting in a trophic upsurge from the increased amount of organic matter available to drive productivity up through the food chain (Kimmel and Groeger 1986, Ploskey 1986, O'Brien 1990, Lintermans 2012, Hatton 2016). Increased submerged habitat area, due to inundation of terrestrial environment, has introduced another food source in the form of displaced terrestrial invertebrates (Hatton 2016). Prior to this time Cotter Reservoir had relatively stable water levels and drought inflows that are likely to be associated with relatively low inputs of organic carbon and/or terrestrial dietary items (Blanchet *et al.* 2008, Winemiller *et al.* 2010). The reservoir has been largely full now for nearly six years, and it appears as though the trophic upsurge has passed and food resources have stabilised and body condition of adults reflect that of a stable reservoir environment (similar to baseline phase).

Abundance of juvenile Macquarie perch in Cotter Reservoir has been relatively stable since 2018, following on from some relatively poor years in late filling / early operation of the enlarged Cotter Reservoir. This has been largely due to a strong contingent of 1+ and 2+ year old individuals. The conversion of 0+ individuals into strong 1+ and 2+ individuals suggests the ECR is providing suitable conditions for early survival and growth of Macquarie perch recruits since autumn 2017.

Fyke netting in 2022 revealed lower abundances of young-of-year Macquarie perch compared to that of 2021 and the other bumper year of operation phase 2017. The presence of young-of-year in the reservoir suggests that access to suitable spawning habitat was achieved in the spawning season of 2021, which comes as little surprise as all reservoirs on the Cotter River were full, and flows were operating as unregulated (similar to that of the successful spawning season of 2016). Based on the operational years monitoring data, Macquarie perch young-of-year abundances are highest during years when reservoirs are full and the river is running largely unregulated during the spawning season. The mechanism that underpins the success of spawning during years when the river is flowing unregulated and reservoirs are full/spilling is likely related to two factors. Firstly, strong flow and temperature cues would be present for the reservoir-resident population of adults resulting from 'natural' flow and temperature conditions of over-dam or spilling flows. This is important to cue and coordinate the spawning response of the adults scattered around the reservoir. Secondly, access to the maximum amount of spawning habitat is provided, as instream barriers are drowned out by higher flows. Conversely, too high a flow in the river during spawning and hatching may lead to washout of spawning sites (Tonkin *et al.* 2017).

Young-of-year abundance at the reference site (Kissops Flat) were at their lowest in 2022 since monitoring began. This continued a decrease in relative abundance of this size class since 2016. It is possible that bushfires in the catchment around Yaouk and Adaminaby over the summer of 2019 / 2020 may have impacted on recruitment of Macquarie perch in 2020 through sedimentation (because of run off) or potentially short-term declines in water quality. In 2021 high discharges throughout the spawning and early recruitment time for Macquarie perch in the upper Murrumbidgee catchment are likely to have negatively impacted on young-of-year abundances. Tonkin *et al.* (2017) found that high discharges during spawning season was negatively associated with recruitment strength of Macquarie perch. The decrease in recruitment strength is likely driven by high water velocities washing out and spawning sites and potentially damaging eggs and early larval phase individuals, or displacing them to less desirable habitat.

RECOMMENDATIONS FOR 2022-23 MONITORING PERIOD

Adult population

Current methods for surveying the majority of the adult Macquarie perch population size classes appear to be adequate, although there may be some size-based or behaviour-based bias in capture efficiency. Based on low captures of large adults in gill nets but increased captures by boat electrofishing in 2018 and again in 2022 (compared to previous years), capture efficiency of larger adults (> 350 mm) in gill nets appears to be reduced, compared with smaller adults. This may be associated with a reduced level of effort regarding the larger mesh gill nets (only shallow drop nets used in 5" mesh size prior to 2019). We recommend continuation of the trial of increased effort to capture larger size classes of Macquarie perch which includes all of the following:

- 1) Additional 125 mm 33 meshes deep 'shallow' gill net;
- 2) Deployment of 125 mm 66 meshes deep 'deep' gill net; and
- 3) Extra boat electrofishing effort (night-time boat electrofishing).

Other than this potential bias in capture efficiency in gill netting, the adult population, especially the largest size classes, appears to be adequately protected. No management intervention is recommended for adult Macquarie perch in the ECR.

Juvenile population

The strong presence of 1+ and 2+ Macquarie perch in ECR indicates that conditions in the reservoir were suitable for survival and growth during the early life history of Macquarie perch in 2019 and 2020. At this stage, no management intervention is recommended for juvenile Macquarie perch in Cotter Reservoir.

Young-of-year

Increased fyke net effort has reduced the variability in young-of-year captures between net nights and provides a comparable level of effort to that of baseline monitoring. We recommended continuation of the increased fyke netting effort in Cotter Reservoir.

The annual detection of a cohort of young-of-year since 2017 is heartening after the consecutive recruitment failures in 2014-2016. Continuing fyke net monitoring as the ECR moves further into operational phase (i.e. use for water supply and further fluctuations in water level) is essential to determine whether reservoir Macquarie perch can spawn and recruit during fluctuating and regulated conditions.

Larval monitoring

It is recommended that snorkelling continues as currently undertaken, as it is a well-tested method that can detect even low numbers of larvae in the Cotter catchment (Broadhurst *et al.* 2012a). The inability to use snorkelling in 2021/22 was an aberration related to high and prolonged periods of high flow, and will rarely occur based on previous experience.

QUESTION 2: Has there been a significant change in the abundance and distribution of Macquarie perch in the Cotter River above and below Vanitys Crossing as a result of the filling and operation of the ECR?

Monitoring for this question could not be undertaken in autumn 2022 because of high discharges of Cotter River during the 2022 autumn monitoring period.

QUESTION 3: Have Two-spined blackfish established a reproducing population in the enlarged Cotter Reservoir and are they persisting in the newly inundated section of the Cotter River?

BACKGROUND

Two-spined blackfish have long been absent from Cotter Reservoir (Lintermans 2002, Ebner *et al.* 2008) (thought to be a result of excessive sedimentation smothering potential spawning sites) apart from a small number of individuals detected in 2012, possibly washed down from the river during flooding (Lintermans *et al.* 2013) (Figure 14). However, the species was present in the river reach inundated by the ECR (Ebner *et al.* 2008, Lintermans *et al.* 2013). Inundated habitats around the perimeter of the ECR should provide suitable spawning habitats for the species. The monitoring program will determine whether the species persists in the newly inundated river reach, and subsequently expands to colonise newly inundated habitats around the perimeter of the ECR.

METHODS

Sampling design for Question 3 follows a similar approach to the baseline monitoring program (Lintermans *et al.* 2013). One of the reference reservoirs from the baseline monitoring program (Corin Reservoir) was dropped from the subsequent (filling and operational) monitoring program to minimise costs.

Table 6. Outline of the sampling design for Question 3 of the fish monitoring program.

Feature	Detail
Target species and life history phase	Two-spined blackfish; Adult (>150 mm TL); juveniles (80 – 150 mm) and young-of-year (<80 mm).
Sampling technique/s	Fyke nets (20 set on the first night around the entire perimeter as part of question 1; then the 8 most upstream nets from nights 2 and 3 of the 20 set as part of questions 1), 12 x 1 night in Bendora Reservoir. 10 x Bait traps (with light stick) set in the newly inundated section of the reservoir.
Timing	Conducted annually in late summer- early autumn.
Number / location of sites	3 sites; 1 around the entire ECR, 1 focussed in the newly inundated area and Bendora Reservoir (reference site).
Information to be collected	Number and total length (mm) for all Two-spined blackfish.
Data analysis	Catch-per-unit-effort (CPUE) assessed between years where possible using 95% (Bonferroni corrected) confidence limits.

Sampling targeted adult, juvenile and young-of-year Two-spined blackfish. Individuals were classed as adults if they are > 150 mm TL, juveniles if 80 – 150 mm TL; and young-of-year if <80 mm TL based on results of Lintermans (1998). At the time of sampling (i.e. late summer / early autumn) young-of-

year will be approximately 50 – 79 mm TL based on results of the baseline data collected (Lintermans *et al.* 2013).

Overnight fyke netting (approx. 16 hours soak time) was used to capture Two-spined blackfish. For the reproduction component of the question all 20 nets from the first night of netting for question 1 was used. For the persistence in the inundation zone component, the eight most upstream nets from nights two and three of sampling undertaken as part of question 1 were used. Sampling for this question is undertaken annually in late summer-early autumn (to be comparable with sampling undertaken in the baseline monitoring program). Two sites within the reservoir were monitored, one around the entire ECR (to detect establishment and recruitment in the ECR), one in the newly inundated section of the ECR (upstream of Bracks Hole reach) and one reference site at Bendora Reservoir. Bait traps were not able to be employed in 2014 as it was not possible to get sufficient number of identical traps in time for sampling (same mesh size, shape, entrance size and colour).

Abundance was standardised as fish caught per net hour (represented as CPUE). Due to the predominance of zero catch data across most samples in Cotter Reservoir, formal statistical tests were not feasible for differences between years. Abundance between years was assessed in Bendora by comparing mean (fish per net hour) CPUE using 95% confidence limits (with Bonferroni correction) overlap.

RESULTS

As for 2021, standard monitoring by fyke nets in the ECR in 2022 captured no Two-spined blackfish (Figure 14). Over six years of monitoring, 2012, 2017, 2018, and 2020 were the only years where Two-spined blackfish was captured in Cotter Reservoir downstream of the newly inundated zone. There was no Two-spined blackfish captured in the bait traps set in the Cotter Reservoir in 2022. Relative abundance of Two-spined blackfish has been stable in the reference site over the monitoring period but has been declining since the 1990s (Lintermans 2001, 2005b). Seven Two-spined blackfish were captured in Bendora Reservoir in 2022, ranging from 162 – 268 mm TL.

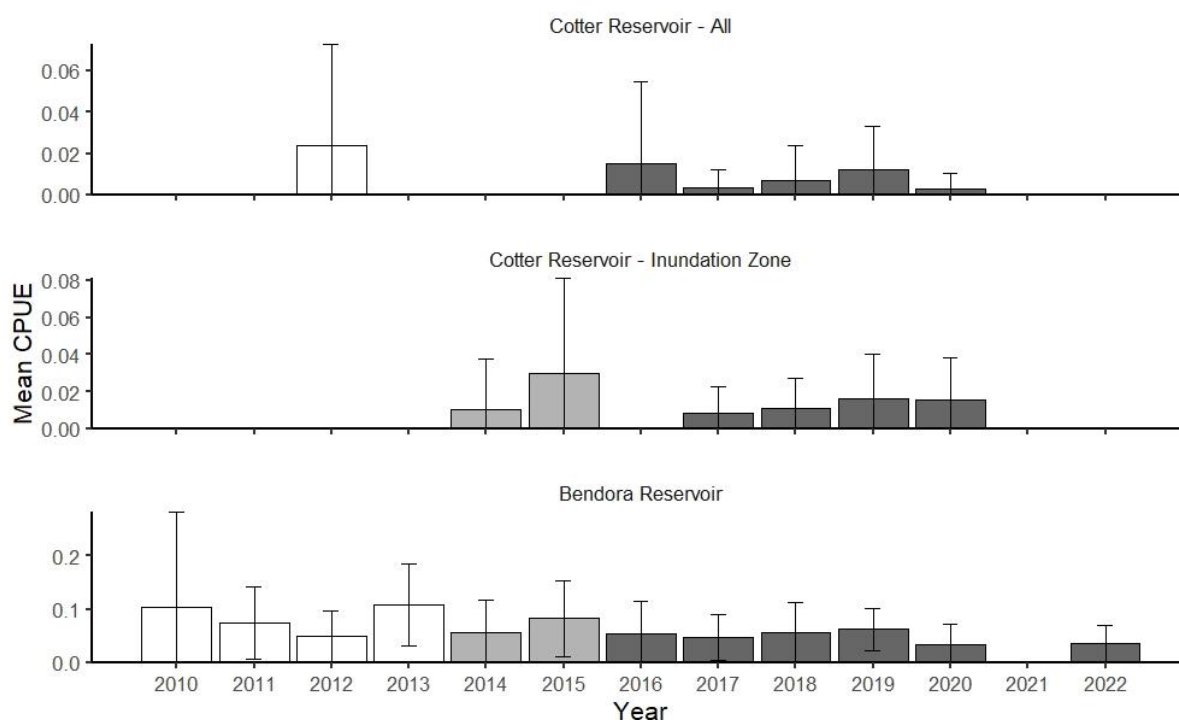


Figure 14. Relative abundance (displayed as mean CPUE \pm 95% confidence limits with Bonferroni correction) of Two-spined blackfish captured by fyke netting in Cotter Reservoir (both all around the reservoir and just the inundation zone) and Bendora Reservoir between 2010 and 2022. For Cotter Reservoir, white bars indicate baseline phase, grey bars indicate filling phase and white bars with diagonal stripes indicates operational phase of monitoring program.

DISCUSSION AND CONCLUSIONS

Two-spined blackfish have been in very low densities in Cotter Reservoir in the 11 years of ECR monitoring. This result supports previous research that identified that the original Cotter Reservoir had sub-optimal habitat for Two-spined blackfish as a result of forestry and associated sedimentation of the reservoir smothering rocky substrate preferred by this species (Lintermans 1998, Ebner and Lintermans 2007, Ebner *et al.* 2008, Broadhurst *et al.* 2011, Broadhurst *et al.* 2012b). The individuals captured in 2012 were at small experimental reefs associated with the *Constructed Homes* project (see Lintermans *et al.* 2010). As previously noted, fyke net sampling of the ECR in April 2016 for the Macquarie perch translocation project, one blackfish was caught on one night and another two were captured on a subsequent night (Lintermans 2017). All individuals were large adults and are not considered to be recaptures (Lintermans unpubl. data). Two of the individuals were captured adjacent to constructed rock reefs on Pryors Road. Translocation sampling with fyke nets in 2019 also captured a large (267 mm TL) blackfish in the non-inundation zone of the reservoir adjacent to constructed rock reefs (Lintermans unpubl. data). In light of this, the large-scale rock reef deployment for adult Macquarie perch has the potential to provide suitable habitat for colonisation by Two-spined blackfish. Monitoring since filling indicates that this is yet to occur on a significant scale and to date no evidence of breeding has been detected in Cotter Reservoir.

The capture of Two-spined blackfish in the newly inundated upstream third of the enlarged Cotter Reservoir in previous years (2018 – 2020) suggest that newly inundated shoreline of the enlarged Cotter Reservoir may serve as suitable habitat for the species, though at low densities. No recruitment of Two-spined blackfish has yet been detected in the ECR. Monitoring over the coming years will provide further clarification of the reservoir's suitability longer-term.

The failure to catch Two-spined blackfish in Bendora Reservoir in 2021 was the first time in 30 years of sampling this reservoir with fyke nets that the species has not been captured (Lintermans unpubl. data). Although Two-spined blackfish were captured in Bendora Reservoir in 2022, relative abundance was quite low, and the status of this population still requires further investigation.

RECOMMENDATIONS

Methods for assessing the population of Two-spined blackfish appear to be adequate. No change to monitoring recommended.

Captures of Two-spined blackfish in the Cotter Reservoir have been rare to this point. At this stage, no management intervention is recommended for Two-spined blackfish in Cotter Reservoir.

Continued low abundance of Two-spined blackfish in Bendora Reservoir is concerning. If the site continues to return very low abundances of blackfish another reference site will need to be sampled (Corin Reservoir).

QUESTION 4: Has there been a significant change in the abundance, distribution and size composition of adult trout in the enlarged Cotter Reservoir as a result of filling and operation?

BACKGROUND

Trout are a potential threat to Macquarie perch in the Cotter Reservoir due to their potential for significant predation of other fishes (Budy *et al.* 2013). An increased reservoir area and depth, and the inundation of terrestrial vegetation were predicted to drive a trophic upsurge that could increase food and/or habitat resources for the resident trout population to increase in abundance and biomass within the Cotter Reservoir (Lintermans 2012). Increased food resources, thermal refuge habitat (increased depth), and improved habitat quality (increased dissolved oxygen as a result of changed destratification procedures) were expected to result in improved growth (and size) of trout individuals, based on their preferred resource requirements (Budy *et al.* 2013). Monitoring changes in the reservoir trout population is needed to give early warning of potential increases in predatory interactions with Macquarie perch.

METHODS

Sampling design for Question 4 is similar to the baseline monitoring program for Question 3 (Lintermans *et al.* 2013) (Table 7). One of the reference reservoirs from the baseline monitoring program (Corin Reservoir) was dropped from the subsequent (filling and operational) monitoring program to minimise costs.

Table 7. Outline of the sampling design for Question 4 of the fish monitoring program.

Feature	Detail
Target species and life history phase	Rainbow and Brown trout; sub-adult and adult fish likely to be piscivorous (> 150 mm FL).
Sampling technique/s	10 Gill nets (fleet of mixed mesh sizes, approx. 6 hours soak time, 5 nights netting in Cotter Reservoir, 2 nights netting in Bendora Reservoir).
Timing	Conducted annually in early autumn.
Number / location of sites	Two sites; enlarged Cotter Reservoir (impact) and Bendora Reservoir (reference), with each site divided into 5 sections.
Information to be collected	Number, location and fork length (mm) for both Rainbow and Brown trout.
Data analysis	Catch-per-unit-effort (CPUE) and adult trout assessed between years (baseline vs. impact), sections and reservoirs using PERMANOVA using the first two nights of netting from each Reservoir. Size of adult trout was compared between years using ANOVA.

Sampling targeted sub-adult and adult Rainbow and Brown trout of a size considered to be piscivorous (individuals of >150 mm Fork Length, FL) with sufficient gape to ingest larval or early juvenile Macquarie perch and Two-spined blackfish (Ebner *et al.* 2007).

Gill netting (as covered in Question 1) was employed to capture trout species, with the exception of the two additional 125 mm gill nets deployed in 2022 that were excluded from analysis for this question. Sampling for this question is undertaken annually in early autumn (so as to be comparable with sampling undertaken in the baseline monitoring program). Two sites were assessed, the impact site (ECR) and a reference site (Bendora Reservoir).

Only Rainbow trout was used in the comparative analyses, as Brown trout does not occur in Bendora Reservoir. CPUE was then scaled to shoreline length at the time of sampling. This was done by multiplying the CPUE for each net night by the proportional change in shoreline as the reservoir filled for a given year. CPUE of trout was compared using a multivariate Permutational analysis of variance (PERMANOVA) in a repeated measures design (highest interaction terms excluded from model) following Anderson *et al.* (2008). Data was $\text{Log}_{10}(x+1)$ transformed then a resemblance matrix was constructed with modified Gower (base 2) dissimilarity measure. Reservoir and phase were treated as fixed factors, and section nested within reservoir and year nested within phase were treated as random factors. Tests were run with 9999 permutations of residuals under a reduced model with Type III sum of squares. Graphical presentations of mean CPUE within each reservoir section (five in total), with 95% confidence limits (with Bonferroni corrections), were used to explore pairwise differences in trout abundance. Size (fork length) variation between years was explored using non-parametric Kruskal-Wallis ANOVA due to severe violations of the data (principally kurtosis) that could be not rectified by data transformation. Summary analysis of the Brown trout CPUE and lengths is also provided for Cotter Reservoir (as they are not present in Bendora Reservoir).

RESULTS

Abundance and distribution

Ten trout were captured in the ECR in 2022, comprising eight Rainbow trout and two Brown trout. This was much lower than the previous two years of monitoring, and similar to that of 2019 (Figure 15). Twenty-four Rainbow trout were captured in Bendora Reservoir in 2022 (Brown trout are not present in this reservoir). There was no significant effect of reservoir, phase or year on the relative abundance of Rainbow trout captured in the ECR (Table 8), though there was a significant effect of section and reservoir by year interaction (Figure 15). The latter was likely driven by the scarcity of Rainbow trout in Bendora Reservoir in 2016 and low abundances again in 2017. The number of Brown trout ($n = 2$) captured in the ECR in 2022 was the second lowest since monitoring began and was by far the lowest number captured since entering operational phase of the ECR.

Table 8. Results of PERMANOVA analysis of gill net catch-per-unit-effort (scaled to relative net effort versus shoreline length at the time of sampling) of Rainbow trout captured in Cotter Reservoir and Bendora Reservoir from 2010 – 2022 (bolded text indicates statistically significant difference at the P(permanova) 0.05 level).

Source	df	SS	MS	Pseudo-F	P(permanova)	perms
Reservoir	1	0.04310	0.0443101	0.68444	0.4295	9831
Phase	2	0.04526	0.0426312	0.9545	0.4088	9955
Section(Reservoir)	8	0.30062	0.0437577	2.2273	0.0266	9946
Year (phase)	10	0.24212	0.0424212	1.4351	0.1593	9932
Reservoir x Phase	2	0.14754	0.0473769	1.2753	0.1131	9909
Reservoir x Year(phase)	10	0.50045	0.0450045	2.9663	0.001	9922
Phase x Section(Reservoir)	16	0.33648	0.042103	1.2465	0.2234	9923
Residuals	470	7.9295	0.0416871			
Total	519	9.7574				

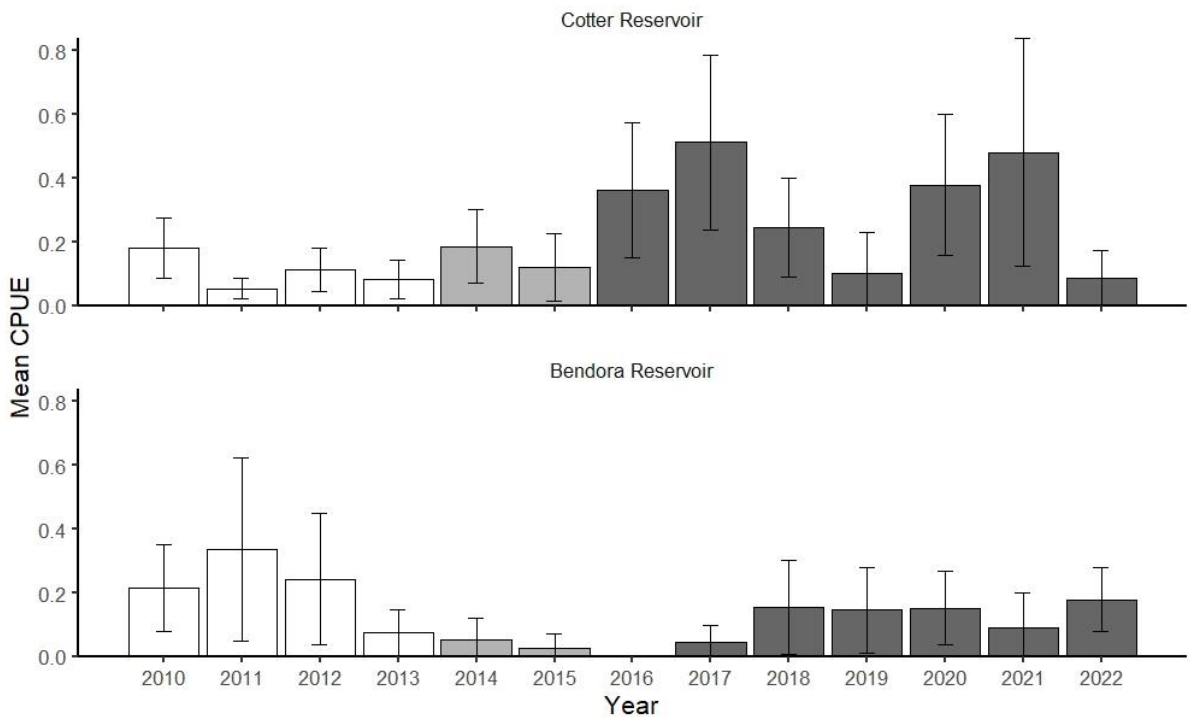


Figure 15. Mean catch-per-unit-effort (\pm 95% confidence limits with Bonferroni correction, scaled for relative net effort versus shoreline length at the time of sampling) of adult Rainbow trout captured in Cotter Reservoir and Bendora Reservoir using gill nets each year from 2010 until 2022. White bars indicate baseline phase, light-grey bars indicate filling phase and dark-grey bars indicates operational phase of monitoring program.

Size composition

Size composition of captured Rainbow trout in Cotter Reservoir has been stable since monitoring commenced. Size of adult Rainbow trout captured in the ECR during 2022 ranged from 330 – 460 mm Fork Length (FL) (Figure 16). Median size of adult trout in the ECR was similar between all years (Figure 18). Size of adult Rainbow trout captured in Bendora Reservoir during 2022 ranged from 244 – 479 mm Fork Length (FL) (**Figure 17**). The two Brown trout captured in gill nets in Cotter Reservoir in 2022 were 403 mm and 485 mm (FL)(Figure 19).

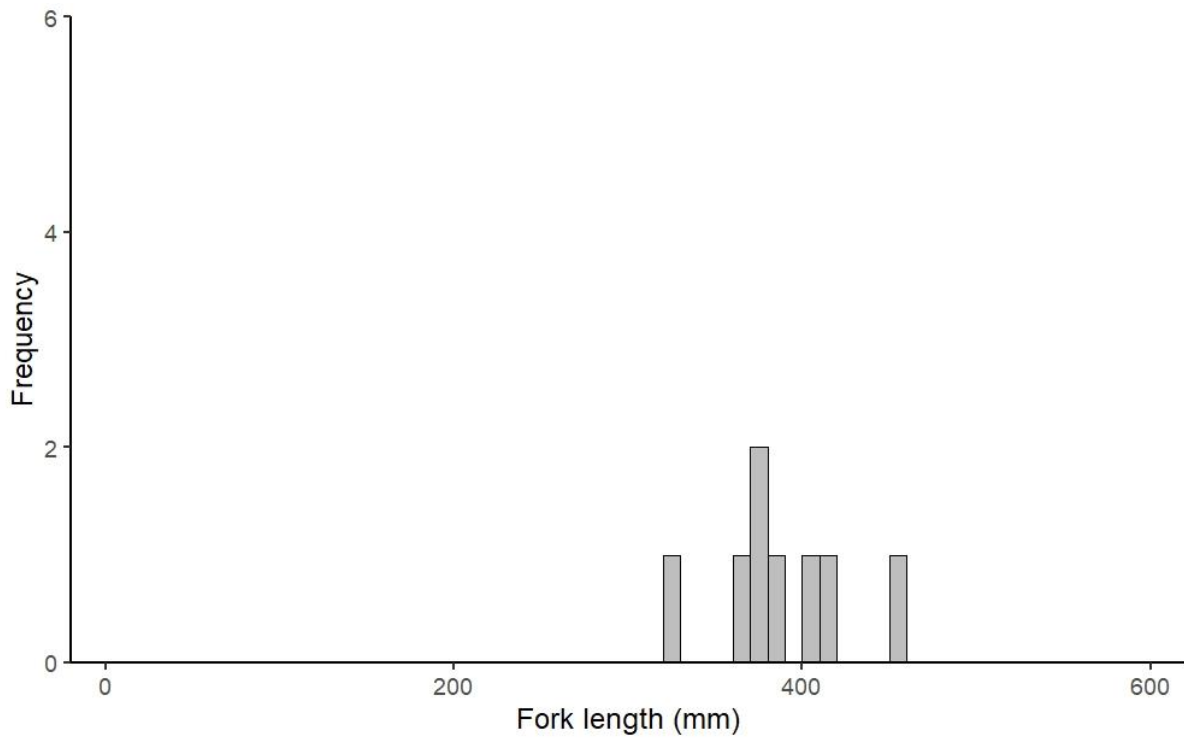


Figure 16. Length frequency of Rainbow trout (n = 8) captured from the ECR in autumn 2022 using gill nets.

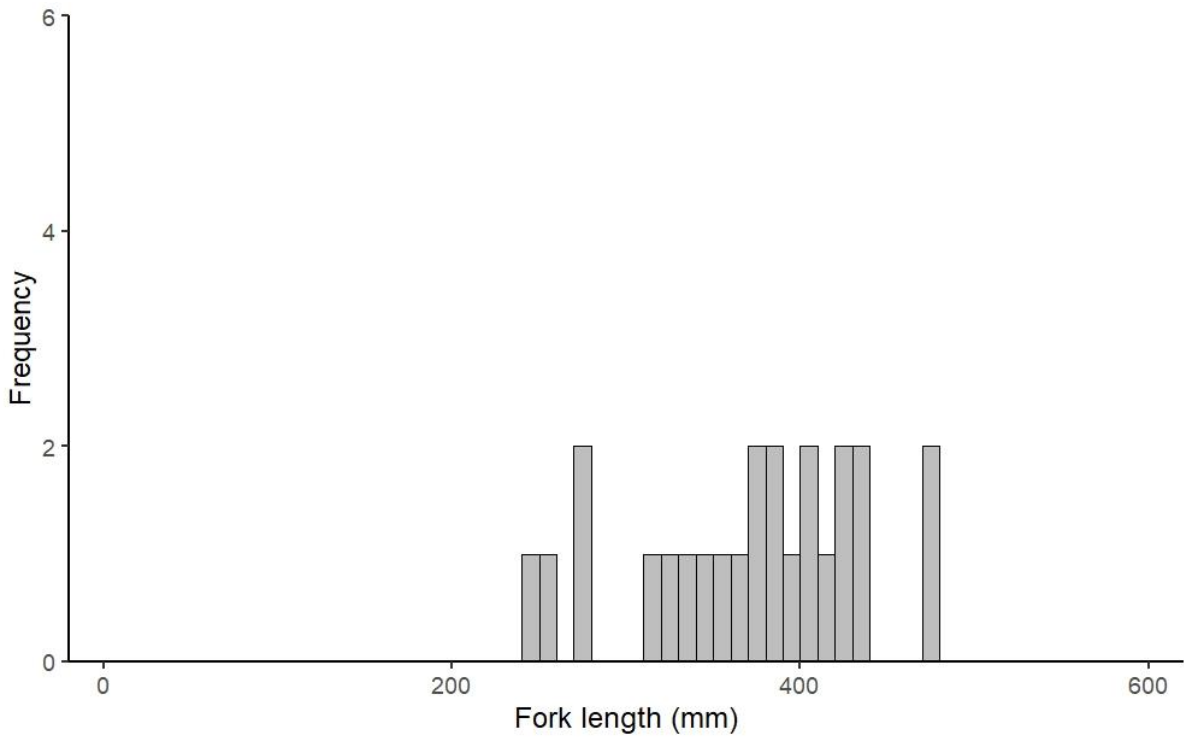


Figure 17. Length frequency of Rainbow trout (n = 24) captured from Bendora Reservoir in autumn 2022 using gill nets.

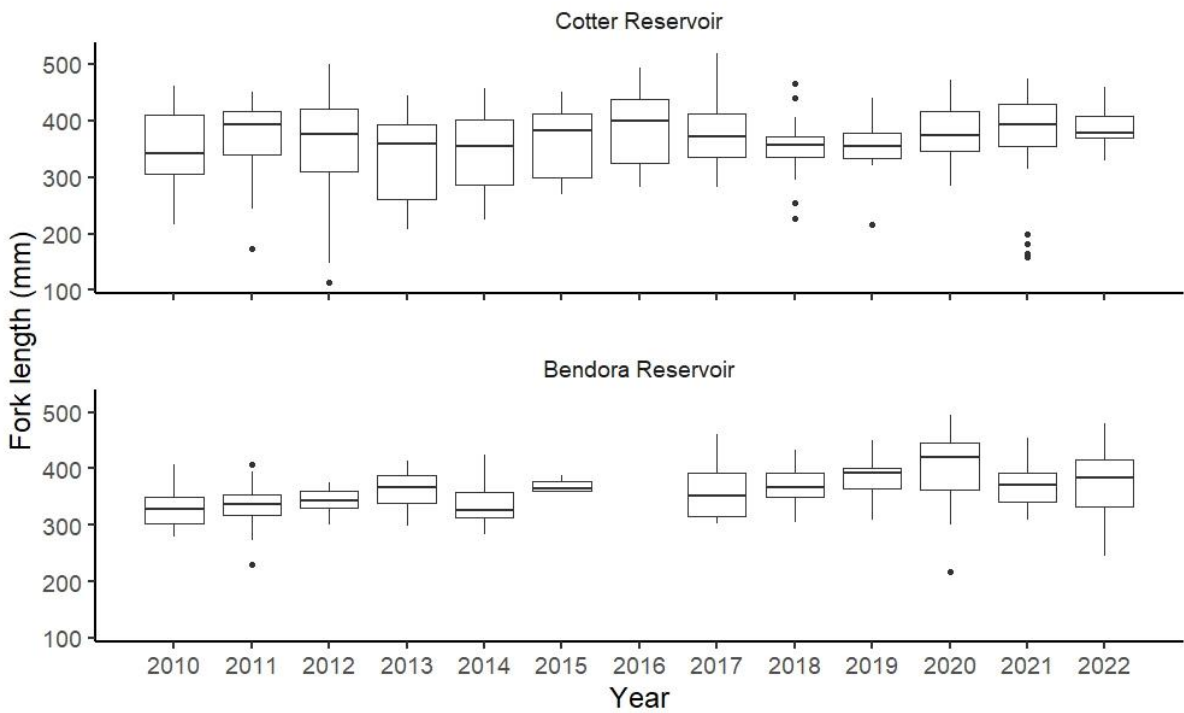


Figure 18. Boxplots of adult Rainbow trout length captured in gill nets each year from Cotter and Bendora Reservoirs from 2010 to 2022 (solid line = median, box represents 25 – 75th percentiles, bars represent minimum and maximum lengths and black circles represent outliers).

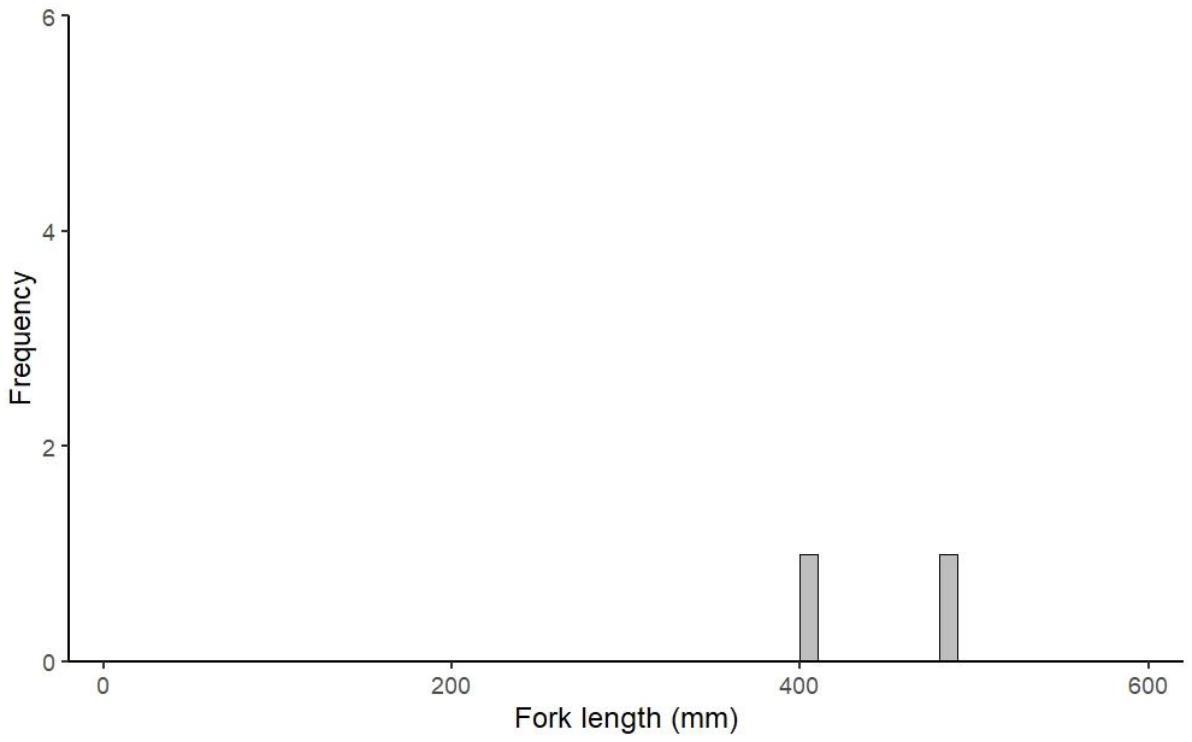


Figure 19. Length frequency of Brown trout (n = 2) captured from the ECR in autumn 2022 using gill nets.

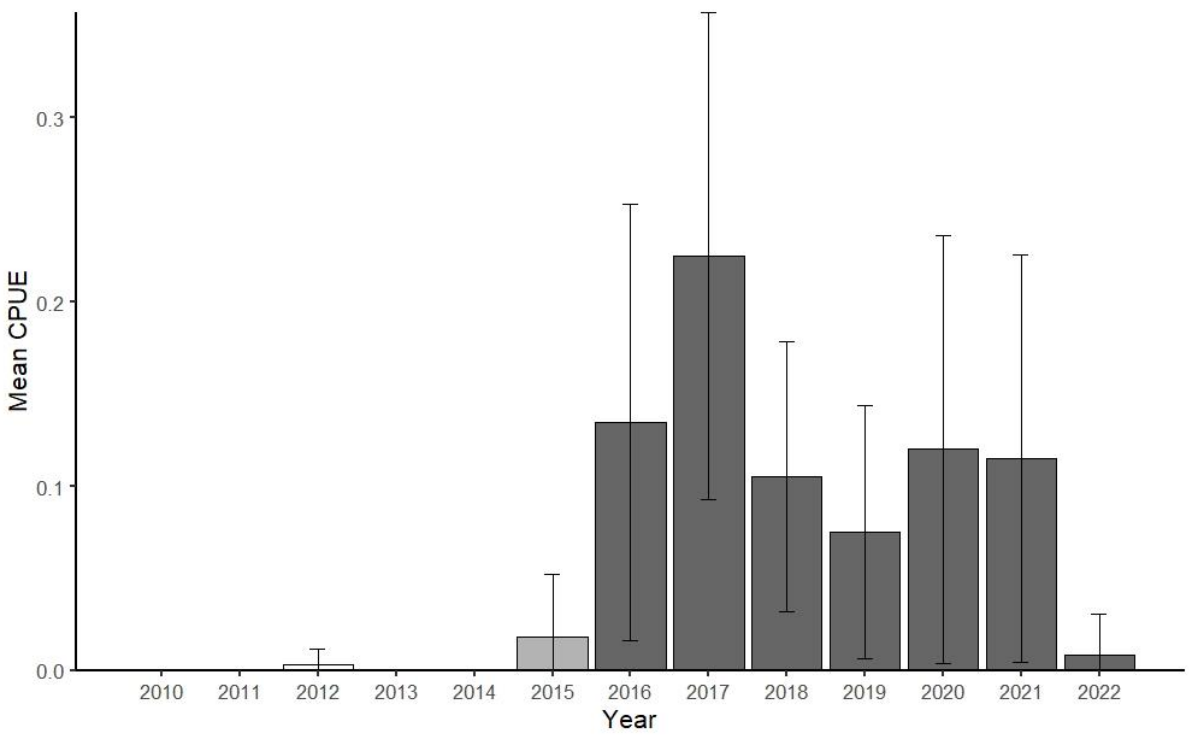


Figure 20. Mean catch-per-unit-effort (\pm 95% confidence limits with Bonferroni correction, scaled for relative net effort versus shoreline length at the time of sampling) of adult Brown trout captured in Cotter Reservoir using gill nets each year from 2010 until 2022. White bars indicate baseline phase, light-grey bars indicate filling phase and dark-grey bars indicates operational phase of monitoring program.

DISCUSSION AND CONCLUSIONS

Abundances of Rainbow trout within the Cotter Reservoir have been somewhat variable over the past 13 years, with higher abundances observed during filling and operational years (especially years of 2016, 2017, 2020 and 2021). The abundance of Rainbow trout in Bendora Reservoir was highest in early baseline years, and has been relatively stable since 2018. Capture of Rainbow trout in Bendora appears to be related to temperature at the time of sampling, where a negative correlation between surface water temperature and number of Rainbow trout captured exists (Table 9). Lowest catches seem to occur when surface water temperature is > 17 °C, with the exception of 2022, which recorded greater than median number of Rainbow trout, even though water temperature was ~20 °C (Table 9).

Table 9. Details of Rainbow trout captures by gill nets and associated surface water temperatures in Bendora Reservoir 2010 – 2022. NR indicates temperature was not recorded.

Year	Date sampled	Water Temperature	No. Rainbow trout
2022	30/3/22; 31/3/22	20.1	24
2021	20/4/21; 21/4/21	15.2	11
2020	20/5/20; 21/5/20	12.5°C	21
2019	29/4/19; 30/4/19	16.2°C	20
2018	5/4/18; 19/4/18	15.0°C	22
2017	1/5/17; 4/5/17	14.2°C	6
2016	29/2/16; 7/3/16	23.7°C	0
2015	10/3/15; 16/3/15	19.7°C	3
2014	13/3/14; 17/3/14	NR	7
2013	4/4/13; 8/4/13	18.3°C	11
2012	19/4/12; 30/4/12	14.9°C	33
2011	9/5/11; 11/5/11	11.2°C	44
2010	20/5/10; 27/5/10	NR	28

During boat electrofishing in Cotter Reservoir, 633 Rainbow trout were observed, with the vast majority of these (79%) being observed during night sampling. The vast majority of these individuals were visually estimated to be sub-adult (<150 mm FL), likely 0 + or 1+ years old. These fish are too small to be captured in gill nets in 2022, but we would expect that in the coming 1 – 2 years, this strong cohort will result in a peak in adult abundance in Cotter Reservoir.

The number of Brown trout captured in the ECR in 2022 was considerably lower than the preceding six years (which had recorded high abundances of Brown trout). Whether the abundance of Brown trout has reduced, or the low captures of 2022 are a relic of sampling or behaviour bias, will become clearer over the next few years. Brown trout remain a threat to Macquarie perch. Anecdotally, Brown trout are considered more piscivorous and potentially more damaging to threatened fish populations than Rainbow trout (NSW Fisheries 2003).

To date, there was no significant change in the size composition of adult Rainbow trout captured between years in Cotter Reservoir. It was expected that as the reservoir filled, food resources would

increase and would lead to increases in size of adult trout. It was considered there was likely to be a time-lag before we see any increases in body length in the resident trout population. Such a change in length would be more likely to occur over multiple years (2 – 5) as the trout would likely first increase their body condition by taking advantage of increased food resources (Kimmel and Groeger 1986, Ploskey 1986, O'Brien 1990), which would over time result in increased growth and length of adult trout. Visual ad hoc examination of trout body condition indicates it has improved post- filling. An increase in the average length of trout (and the maximum length) will likely change predation dynamics in the reservoir as larger trout (with larger gape-range) seek larger food items that tend to be resident prey fishes (Jonsson *et al.* 1999, Ebner *et al.* 2007). So far, this has not materialised, though a scenario of considerable conservation concern would be the growth of extremely large trout that have the capacity to consume a wide range of sizes of Macquarie perch. It remains to be seen what will happen to length (and body condition) if the observed large numbers of juvenile trout persist to adulthood.

RECOMMENDATIONS

Methods for assessing the population metrics of adult trout relative abundance, distribution and size appear to be adequate. No changes to monitoring are recommended.

No management response to the Rainbow trout in Cotter Reservoir is recommended at this time. However, it is still considered a risk that trout size and abundance may increase over time, and modelling has shown that trout predation can have significant impacts on blackfish in the Cotter River (Todd *et al.* 2017). These potential impacts indicate that there is still value in investigating potential trout control mechanisms so that management action could be deployed should an increased abundance or size be detected in subsequent monitoring (Lintermans 2012, ACTEW Corporation 2013). The recovery of the Brown trout population in the Cotter catchment from 2016 – 2021 is of some concern, with this species being highly piscivorous once they attain length of > 250 mm FL (all of the 2017 – 2020 and 2022 and most of the 2021 captures were above this size). The increased abundance of Brown trout (and particularly large brown trout) needs to be monitored closely and appropriate management action taken if the current trend continues. No management of this species is currently warranted, but a more rigorous investigation of the diet of Brown trout should be considered should abundance rebound from the low captures of 2022 in the coming years.

QUESTION 5: Has there been a significant change in the abundance and size composition of trout in the Cotter River upstream of the enlarged Cotter Reservoir as a result of filling and operation?

Monitoring for this question could not be undertaken in autumn 2022 because of high discharges of Cotter River during the monitoring period

QUESTION 6: Are Two-spined blackfish and Macquarie perch present in trout stomachs in the Cotter River?

Monitoring for this question could not be undertaken in autumn 2022 because of high discharges of Cotter River during the monitoring period

QUESTION 7: Has there been a significant change in the abundance and distribution of non-native fish species in the enlarged Cotter Reservoir as a result of filling and operation?

BACKGROUND

The dynamics of the trout population in the ECR is addressed by Question 4. The other non-native species present in Cotter Reservoir are Goldfish *Carassius auratus*, Oriental weatherloach *Misgurnus anguillicaudatus* and Eastern gambusia *Gambusia holbrooki*, all of which have noted preferences for still-water or slow-flowing habitats (Lintermans 2002, 2007). The enlargement of the reservoir provided a significant increase in habitat for these species as well as the trophic upsurge, and consequent increases in abundance were observed in the first few years since filling commenced. These species could competitively interact with Macquarie perch for resources (particularly food and shelter) but are not considered a predatory threat. Increased Gambusia abundance could lead to increased aggressive interactions between this and native fish species (Lintermans 2007). Also, expansion of populations of Goldfish and Oriental weatherloach could facilitate the expansion of trout and cormorant populations, which are a potential predation threat to threatened fish populations. Both Goldfish and Oriental weatherloach have been recorded in trout diet from the reservoir, with Goldfish being particularly important (Ebner *et al.* 2007). Cormorant diet in the Cotter Reservoir has also been shown to contain significant numbers of Goldfish (Lintermans *et al.* 2011). Monitoring changes in status of non-native fish in the reservoir, along with monitoring of trout predation in the river (Question 6) will provide insights into the dynamics of the fish community in the reservoir. This monitoring will also facilitate early detection for non-native fish species not currently in the Cotter catchment upstream of Cotter Dam (i.e. Carp & Redfin perch).

METHODS

Sampling design for Question 7 is covered by sampling outline for Question 1 (fyke netting) and is similar to the baseline monitoring program (Lintermans *et al.* 2013) (Table 10). The changes from the baseline monitoring program are the removal of an urban lake reference site (where these non-native species are present/abundant). This is the seventh year of monitoring following the commencement of filling and the fifth year of monitoring since the ECR filled.

Table 10. Outline of the sampling design for Question 7 of the fish monitoring program.

Feature	Detail
Target species and life history phase	Non-native species (other than trout); all sizes.
Sampling technique/s	Fyke nets (20 per night for 3 nights) and bait traps (10 traps for one night).
Timing	Conducted annually in late summer / early autumn.
Number / location of sites	1 site; ECR.
Information to be collected	Number and total length or fork length (mm) for all species.
Data analysis	Comparison of catch-per-unit-effort (CPUE) of non-native fish species between years using ANOSIM. Graphical representations of the means are provided (with 95% confidence limits with Bonferroni corrections).

Sampling targeted all non-native fish species and life stages (other than trout). Fyke netting was used to monitor Oriental weatherloach and Goldfish. Specifically, 20 fyke nets were set around the entire ECR over three nights. 10 Bait traps were set for one night around the perimeter of the reservoir. Sampling for this question was undertaken in early autumn. Total length (TL) and/or fork length (FL) in mm to be recorded for all captured individuals.

Abundance of Goldfish was standardised for each technique as fish caught per hour (represented as catch per unit effort or CPUE). CPUE was scaled in relation to increases in shoreline length as the reservoir filled and as net effort (see question 1 for scaling equation). CPUE of Goldfish captured in fyke nets was assessed between years using analysis of similarity (ANOSIM) with year as fixed factor. Data was $\text{Log}_{10}(x+1)$ transformed then a resemblance matrix was constructed with modified Gower (base 2) dissimilarity measure transformed to meet the assumptions of sphericity and homoscedascity of variances. Graphical presentations of site-level means with 95% confidence limits (with Bonferroni corrections applied) were then used to explore pairwise variations in Macquarie perch size classes among sites and years.

RESULTS

Goldfish

Over three nights of fyke netting in Cotter Reservoir in 2022, 58 Goldfish were captured ranging in length between 36 – 160 mm FL (Figure 21). The vast majority of these individuals were less than 100 mm FL, most likely corresponding to 0+ and 1+ year-old age class (Merrick and Schmida 1984)(Figure 21). Goldfish relative abundance was significantly different among years (Global $R = 0.122$, $p < 0.001$), with relative abundance in filling years of 2014 and 2015 and operational year 2016 significantly higher than in baseline years 2010, 2012 and 2013 and the most recent operating phase years (2017 – 2022) (Figure 22). Relative abundance of Goldfish from 2017 – 2022 was not significantly different from any of the baseline monitoring years.

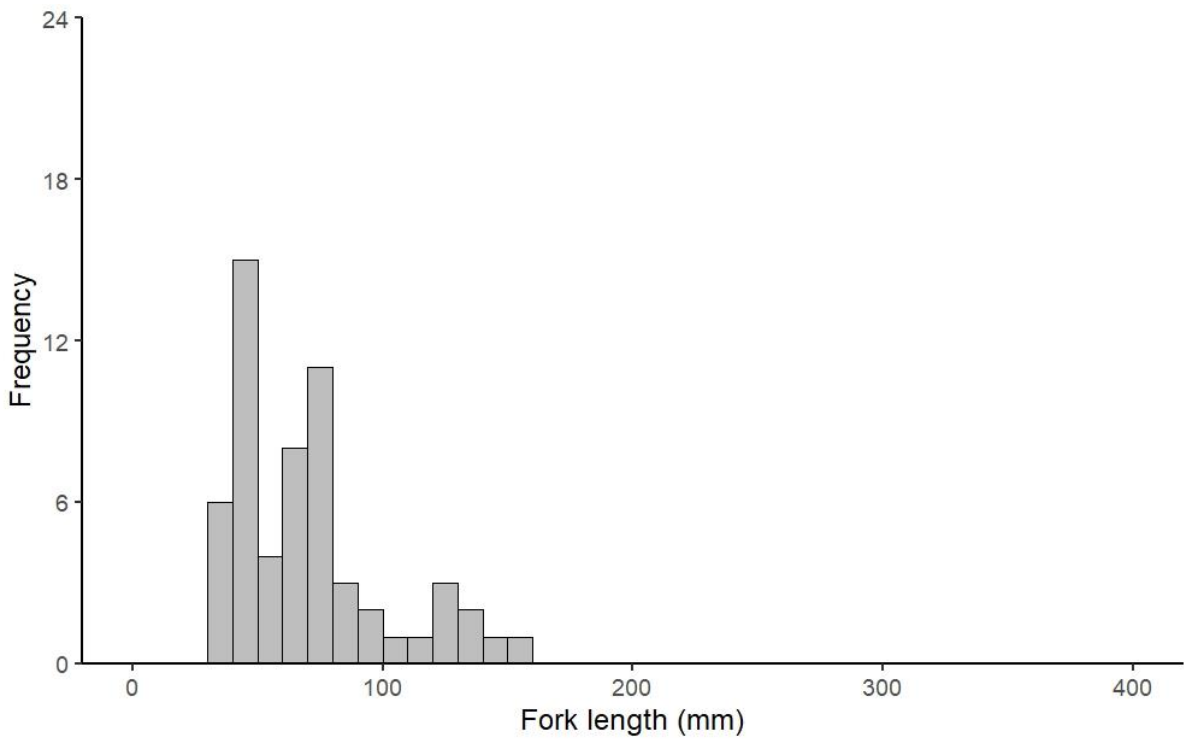


Figure 21. Length Frequency of Goldfish captured in the ECR in 2022 over three nights of fyke netting.

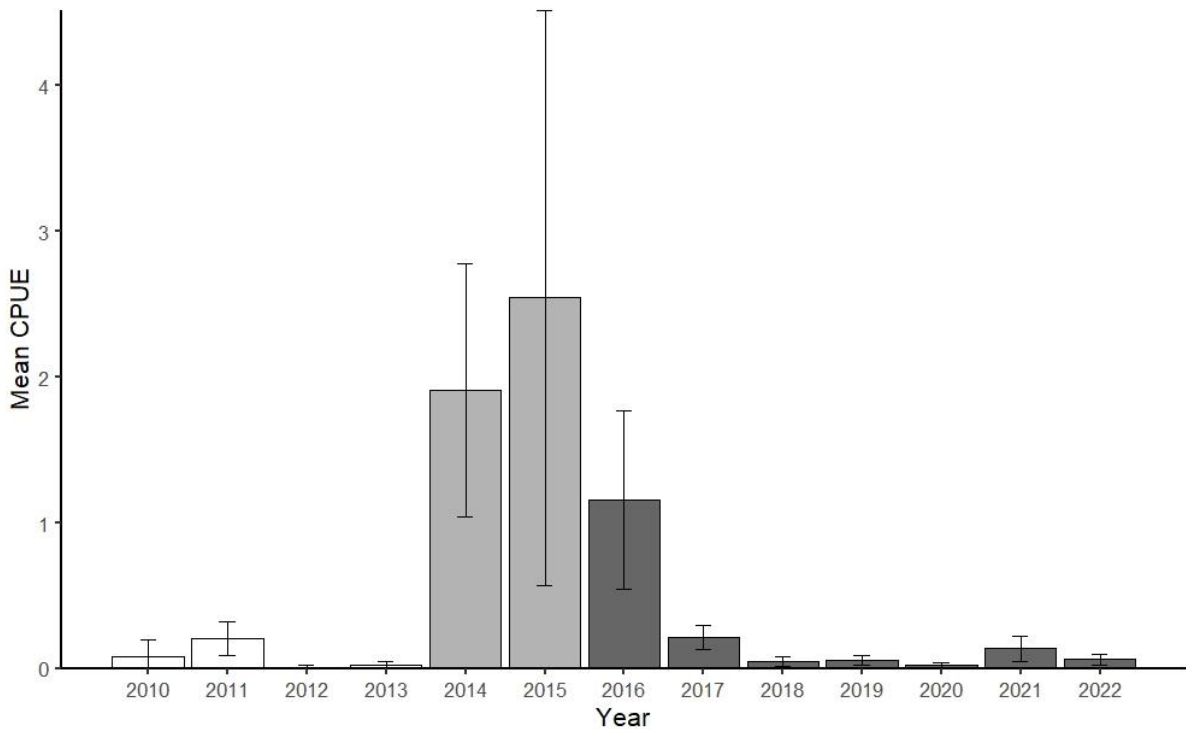


Figure 22. Relative abundance (displayed as mean CPUE \pm 95% confidence limits with Bonferroni correction, scaled for relative net effort versus shoreline length at the time of sampling) of Goldfish captured in the ECR using fyke nets between 2010 and 2022. White bars indicate baseline phase, light grey bars indicate filling phase and dark grey indicates operational phase of monitoring program.

Oriental weatherloach and Eastern gambusia

Both Oriental weatherloach and Eastern gambusia are rare catches in Cotter Reservoir in the monitoring undertaken to date. A total of seven Oriental weatherloach have been captured so far in ten years of monitoring in Cotter Reservoir (one each in 2010 and 2011; three in 2012 and two in 2017). Thirty-seven Oriental weatherloach have been observed whilst undertaking the boat electrofishing of Cotter Reservoir (30 in 2016 and four in 2017). No Oriental weatherloach were observed or captured in 2022 via any of the sampling methods.

Eastern gambusia have only been captured in fyke nets in three years, 2014, where 13 individuals were captured, all from one fyke net and 2015 and 2017 where one individual was captured, respectively. Eight Eastern gambusia (ranging from 18 – 33 mm TL) were captured in bait traps in 2015 and a further seven captured in bait traps in 2016. A single eastern gambusia was captured in bait traps in 2021 (22 mm TL). No fish were captured in bait traps in 2017 – 2020. It must be noted that large numbers of Eastern gambusia were observed whilst undertaking the boat electrofishing during three years of monitoring (2014 – 2016). However, only 29 were observed while boat electrofishing in 2017 and only 1 observed in 2018. One Eastern gambusia was observed during the night in 2022. Small numbers of Eastern gambusia are also regularly observed in the reservoir at night during gill netting operations.

DISCUSSION AND CONCLUSIONS

Relative abundance of Goldfish has been relatively low since 2017, following on from very high abundances during filling and the first year of operational phase. The higher relative abundance of Goldfish in the initial stages of filling was likely to have been caused by an increase in availability of food resources associated with the filling phase (and refilling in the case of 2021) of the ECR (Kimmel and Groeger 1986, Ploskey 1986, O'Brien 1990). As predicted, the increase in Goldfish observed in 2014 – 2016 was likely a significant factor to the commencement of cormorant breeding observed in the top end of the reservoir in February 2014 and in every year since (see Question 8). Goldfish abundances in operational years of 2017 – 2022 were not significantly different to baseline abundances, indicating that the boom in resources associated with filling and early operational phases has ceased.

The relatively low catch rates again of both Oriental weatherloach and Eastern gambusia is not considered to reflect population sizes of these two species, especially that of Eastern gambusia. The low capture rates are likely to be an artefact of the sampling method (fyke nets and bait traps) and the size or behaviour of (Eastern gambusia) or body-shape (narrow cylindrical, Oriental weatherloach) of these two species. The mesh size of the fyke nets used is too large to reliably capture either species. Eastern gambusia is regularly observed in large schools (sometime in excess of 100 individuals) at the boat ramp and other open shallow habitat (such as where existing roads run into the reservoir) at Cotter Reservoir during monitoring. Eastern gambusia are known to prefer shallow waters (Pyke 2005, Lintermans 2007, Macdonald and Tonkin 2008) which may explain the congregations at these shallow open habitats. Certainly, many of these congregations were observed in 2015 and 2016 during the boat electrofishing surveys, but not as many in 2017 or in 2018. The lack of adequate representation of Oriental weatherloach and Gambusia populations in the monitoring program is not of major concern, as the major target of this research question is Goldfish and their likely role in the expanding predator (trout and cormorant) populations.

RECOMMENDATIONS

Sampling methods appear to be adequate for monitoring abundances of Goldfish species in Cotter Reservoir. No change to monitoring regime for this species recommended at this time.

Both Oriental weatherloach and Eastern gambusia have low catch rates in the ECR monitoring program, and observation of localised schools of Eastern gambusia around shallow exposed areas of the reservoir indicate that this species can be numerous at a small spatial scale. Alternative sampling techniques are available for these two species (e.g. seine netting shallow habitats for Gambusia; backpack electrofishing of soft substrates for Oriental weatherloach), or eDNA for both species, but such sampling will require additional sampling days, for data currently considered to be of little consequence for management of threatened fish species. No management intervention required for these two species.

A significant increase in the abundance of Goldfish was detected in 2014 and 2015 and higher relative abundances persisted into 2016. Monitoring from 2017 – 2022 revealed a decrease in Goldfish abundances indicating that the resources boom associated with the filling reservoir has ceased or slowed significantly. The decline in Goldfish abundance and the first detections of Macquarie perch predation by trout warrant close attention being paid to this potential shift in food webs, but at this stage, no management intervention specifically related to Goldfish is recommended.

QUESTION 8: Has there been a significant change in the abundance, distribution and species composition of piscivorous birds in the vicinity of the enlarged Cotter Reservoir as a result of filling and operation?

BACKGROUND

Piscivorous birds (predominantly cormorants) have been identified as a potential threat to Macquarie perch in the ECR (Lintermans 2005a). Predation of Macquarie perch by cormorants in Cotter Reservoir has been confirmed (Ebner and Lintermans 2007, Lintermans *et al.* 2011, Lintermans 2012), and a significant expansion of the piscivorous bird population following enlargement of the reservoir could have severe consequences on the small adult population size of Macquarie perch (Farrington *et al.* 2014). Assessment of population trend in piscivorous birds on Cotter Reservoir is required with monthly monitoring enabling early detection of significant changes in the abundance and distribution of cormorant species. A cormorant management plan has been included in the fish management plan version 4 (Icon Water Limited 2019).

METHODS

Sampling design followed that exactly outlined in the baseline monitoring program (Lintermans *et al.* 2013) (Table 11).

Table 11. Outline of the proposed sampling design for Question 8 of the fish monitoring program.

Feature	Detail
Target species and life history phase	Piscivorous bird species (incl. Great cormorants, Little black cormorants and Little pied cormorants, Darters and Pied cormorants).
Sampling technique/s	Visual survey of piscivorous birds per section (longitudinal fifth) of the ECR.
Timing	Monthly, year-round.
Number / location of sites	1 site; ECR.
Information to be collected	Species, abundance, abundance per section.
Data analysis	Comparison of abundance and distribution of each species of cormorants between years (baseline vs. impact) using Multivariate analysis PERMANOVA. Graphical representations of the means are provided (with 95% confidence limits with Bonferroni corrections).

Monthly visual surveys are undertaken of the entire ECR targeting piscivorous bird species including Great cormorant, Little black cormorant, Little pied cormorant, Pied cormorant, and Darter. The presence of nests of piscivorous birds was also noted, and if present the contents (eggs or chicks) noted (though this was not part of the monitoring program or analysis). Visual surveys were conducted from a boat using 10 x 40 mm binoculars. Location of each individual was recorded on a

map. To determine distribution of piscivorous birds, the reservoir was divided longitudinally into five equal parts. Abundance and distribution can be assessed against trigger levels in the fish management plan: Appendix G (Icon Water Limited 2019). Comparison of abundance and distribution of each species of cormorants between the three phases (baseline, filling, operational) is undertaken using multivariate analysis (PERMANOVA) to explore overarching structure in the cormorant community. Unbalanced permutational analysis of variance (PERMANOVA) was conducted on cormorant abundances. Data was $\text{Log}_{10}(x+1)$ transformed then a resemblance matrix was constructed with modified Gower (base 2) dissimilarity measure. For PERMANOVA analysis, monitoring phase and section were fixed factors, with year nested within phase (Anderson *et al.* 2008). Highest interaction term removed for repeated measures design. Type III Sum of Squares used to account for unbalanced (years across phase) design and the three species of cormorant used as variables.

RESULTS

Great, Little black and Little pied cormorants were the most abundant species of piscivorous birds recorded on the ECR with much lower numbers of Darter and zero Pied cormorant recorded in 2021-2022 (Figure 23). There have been only six observations of Pied cormorant (all of single individuals) since monitoring began in 2010, though none in 2018, 2019, 2021 and 2022. Abundances of the three most common species were relatively consistent with expectations during the monitoring period with some seasonal fluctuations present (Figure 23). Since filling began, abundances of both Great cormorant and Little black cormorant have been stable, though with some definitive seasonal fluctuations (Figure 23). Abundances of Little pied cormorant during warmer months has been increasing annually since filling began, with these annual influxes concentrating in section 4 and as of 2018 section 2 of the reservoir (Figure 23 and Figure 24).

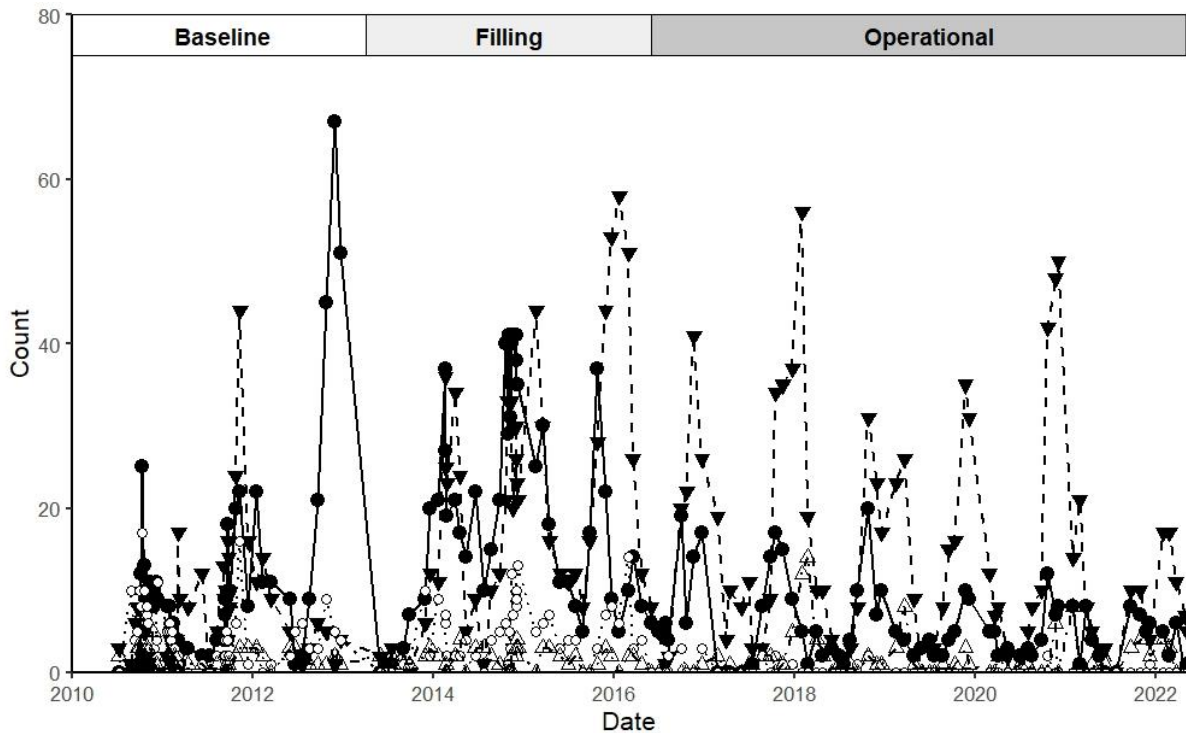


Figure 23. Monthly abundances of each common piscivorous bird species on the Cotter Reservoir between July 2010 and May 2022 (Great cormorant – solid line with black circles; Little black cormorant – dotted line with white circles; Little pied cormorant – dashed line with inverted black triangles; Darter – dash-dot line with white triangles).

There was considerable overlap in the composition of the piscivorous bird community (composition, abundance and distribution) among pre-filling and filling phases in Cotter Reservoir. There was a significant difference among phase, section, year and section by phase interaction in terms of piscivorous bird community composition (Table 12). Pairwise comparisons indicate these significant phase x section interaction differences are among baseline and filling versus operational phase for sections 1 to 4 ($p < 0.05$).

Table 12. Results of PERMANOVA analysis of piscivorous bird community composition in Cotter Reservoir from 2010 – 2022 (bold text indicates statistically significant difference at the P(permutation) 0.05 level).

Source	df	SS	MS	Pseudo-F	P(permutation)	Unique permutations
Phase	2	52.125	26.062	15.206	0.0001	9947
Section	4	42.088	10.522	18.157	0.0001	9929
Year (within Phase)	10	16.825	1.6825	2.9033	0.0001	9899
Section x Phase	8	51.309	6.4136	11.068	0.0001	9910
Residuals	840	486.78	0.5795			
Total	864	660.92				

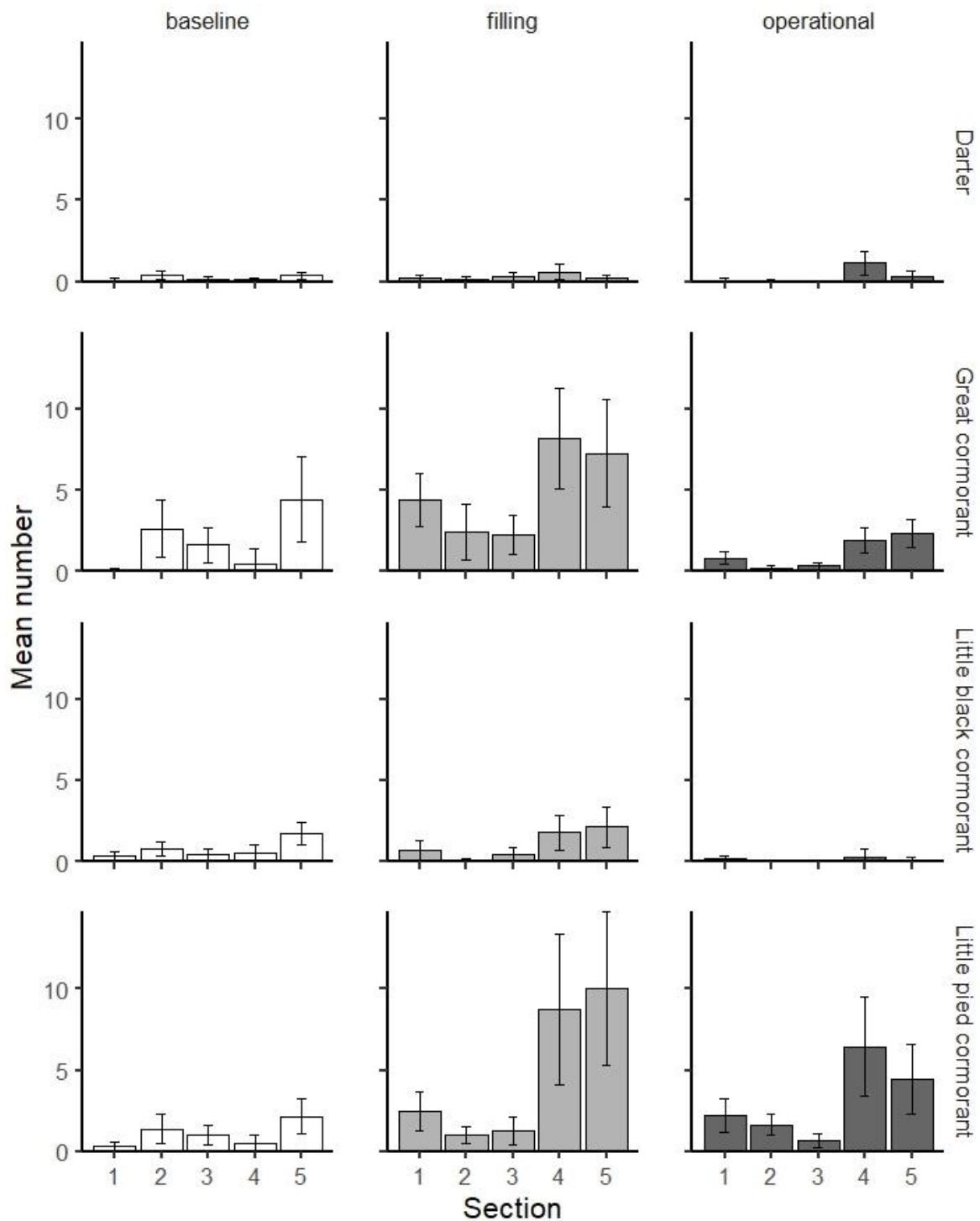


Figure 24. Mean monthly abundance (displayed as mean \pm 95% confidence limits with Bonferroni correction) of each piscivorous bird species in each section of Cotter Reservoir for baseline phase (July 2010 – March 2013), filling phase (April 2013 – December 2015) and operational phase (January 2016 - May 2022) of monitoring program.

For the eighth consecutive year cormorants have established a breeding colony on the ECR. Nesting has occurred in the same reservoir section across years (in sections 4 and 5 of the reservoir approximately 200 m downstream of the Pierces Creek junction Figure 25), though a nesting site was found in 2018 in section 1 (no evidence of nesting here in 2022). Many of the nests observed had chicks varying from just hatched to well-developed. It is believed that the bulk of these were Little pied cormorant as the adult birds were observed on the nests, though the colonies also contained Darters and for the first time confirmed in 2022, a small number of Great cormorant nests.

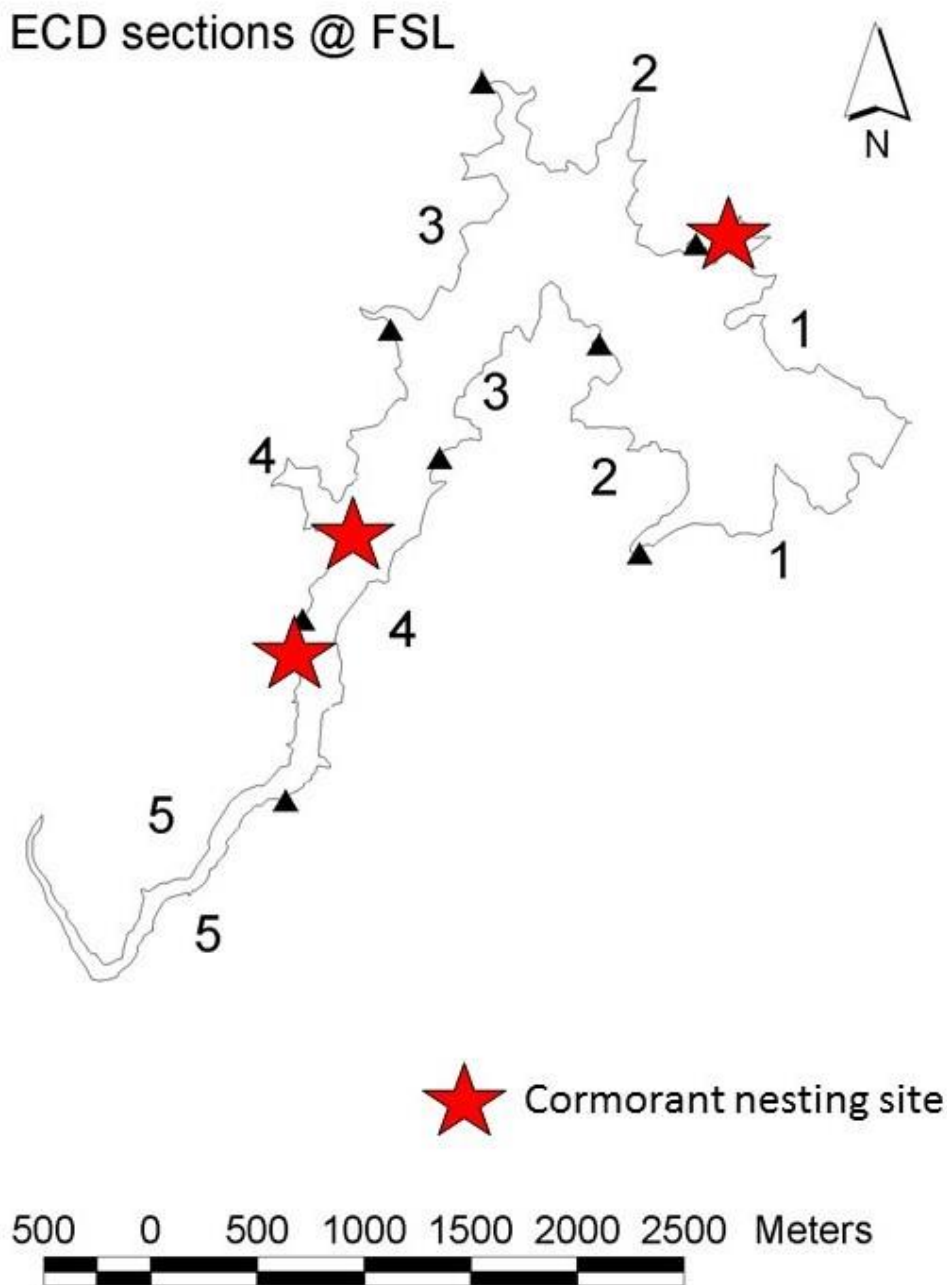


Figure 25. Map of the enlarged Cotter Reservoir shoreline sections with Cormorant nesting colony locations.

DISCUSSION AND CONCLUSIONS

Abundance

Peak abundances (see Figure 29) of Great cormorants occurred during the filling phase, though since the operational phase began have stabilised, though with some definite seasonal fluctuations. Abundances of Little pied cormorant during warmer months have increased since filling began, with these annual influxes concentrating in section 4 of the reservoir. The seasonal increases in all three common species is most likely attributable to an increase in productivity and food resources (decapods and Goldfish) within the reservoir during the warmer months. During filling and early operational phases the enlarged reservoir has seen an increase in the abundance of Goldfish (see Question 7 above), a favoured prey item of cormorants in the Cotter Reservoir and elsewhere (Miller 1979, Lintermans *et al.* 2011). The increase in prey abundance and the abundance of partially inundated larger trees (predominantly Eucalypts and pine trees) has provided suitable conditions for nesting to commence. Indeed, Little pied cormorant has bred in all seven years since filling began and its seasonal abundance peaks during the warmer months increasing until ~2018, though appear to be subsiding each year since ~2019. Cormorants are opportunistic and nomadic, responding to 'boom' conditions, and will breed if resources are sufficient (Kingsford *et al.* 1999, Dorfman and Kingsford 2001b). A 'boom' in food resources in the ECR and the presence of emergent flooded trees has resulted in the establishment of a breeding population of cormorants. The establishment of a breeding colony of cormorants in the ECR is undesirable as the energy requirements of maintaining fledglings as well as adults would incur increased pressure on food resources (i.e. Goldfish and Macquarie perch) by cormorants in Cotter Reservoir (Lintermans *et al.* 2011). The potential early signs of a shift in predation pressure by trout (i.e. first records of predation of Macquarie perch) in [year] potentially associated with declines in Goldfish abundance suggest that a re-examination of cormorant diet is required in the near future. This is especially so as the major avian predation threat to Macquarie perch (Great cormorant) were confirmed as breeding for the first time in 2022. If cormorants are also shifting their food preference from Goldfish to Macquarie perch then management of cormorant breeding colonies becomes critical.

Distribution

Distribution of all three common cormorant species has been relatively stable during baseline (2010 – 2013), filling and operational phases with a few exceptions. All three species have been most abundant in the two upstream sections of the reservoir. Previous research has found that cormorants commonly hunt in depths of less than 5 m (Dorfman and Kingsford 2001a, Ropert-Coudert *et al.* 2006) and this depth range is most prevalent in these two reservoir sections and provides the greatest area for which effective hunting can be conducted (Ryan 2010, Ryan *et al.* 2013). The most upstream section is also where the greatest risk of predation is for Macquarie perch (Ryan 2010, Lintermans *et al.* 2011, Ryan *et al.* 2013). Interestingly a change in the distribution has occurred between baseline and filling and operational phases, where section 4 has seen an increase in abundance of both Great cormorant (during filling only) and Little pied cormorant (both filling and

operational). This is most likely attributable to the location of a nesting colony in section 4 (Figure 25) and associated roost that was not present in baseline monitoring.

RECOMMENDATIONS

The current monitoring regime appears to be adequate at monitoring abundances and distributions of cormorant species in Cotter Reservoir. No changes to the monitoring regime are recommended at this time.

An increase in cormorant abundance and multiple cormorant nesting events have been detected since filling. The increase in abundance of both Great cormorant and Little pied cormorant triggered management action under the ECR Cormorant Management Plan in 2016. The management trigger thresholds in the Cormorant Management Plan have been revised to reflect the likely normal increase in cormorant abundance with an increasing reservoir surface area and shoreline length.

Given the significant decline in the Goldfish population, and the continued presence of breeding colonies of cormorants (including Great cormorant in 2022), it may be that the increased cormorant abundance and presence of breeding colonies may need to be sustained by another fish species (i.e. Macquarie perch). This highlights that a re-examination of cormorant diet is required in the near future. If cormorants are also shifting their food preference from Goldfish to Macquarie perch then management of cormorant breeding colonies becomes critical.

QUESTION 9: Have macrophyte beds re-established in the enlarged Cotter Reservoir?

BACKGROUND

Existing macrophyte beds in Cotter Reservoir have been demonstrated to provide important daytime resting habitat for adult Macquarie perch (Ebner and Lintermans 2007). It was known that existing macrophyte beds would be drowned by up to 50 m of water once the reservoir filled. Modelling indicated that the reservoir would remain within 3 m of full supply level for at least 73 percent of the time once the reservoir has filled, potentially allowing new macrophyte beds to establish. Such macrophyte beds could once again provide important cover habitat for threatened fish including Macquarie perch.

DOES COMPARABLE BASELINE DATA EXIST?

Partially. Surveys by Roberts (2006) and Ryan (2010) provide an indication of macrophyte extent and distribution in the current Cotter Reservoir.

SAMPLING DESIGN

Sampling design will be a standard on-ground survey (e.g. Roberts 2006) of the perimeter of the ECR for signs of establishment of macrophytes. It is considered unlikely macrophytes will establish during filling phase and so the project team recommends that surveys for macrophyte establishment do not commence until ECR has reached full supply level.

Table 13. Outline of the proposed sampling design for Question 9 of the fish monitoring program.

Feature	Detail
Target species and life history phase	The survey will target emergent macrophyte species that are likely to provide adult Macquarie perch with cover from cormorant predation (i.e. <i>Phragmites australis</i>).
Sampling technique/s	Visual on-ground survey of the perimeter of the ECR by boat.
Timing	Conducted monthly during spring and summer.
Number / location of sites	1 site, the entire ECR.
Information to be collected	Location, extent (length / area covered) of each emergent macrophyte species.
Data analysis	Descriptive statistics length, width and area of each species and stand. Map of size and location of macrophyte stands will be derived.

The survey will target emergent macrophyte species that are likely to provide adult Macquarie perch with cover from cormorant predation (i.e. *Phragmites australis*). Visual surveys will be conducted around the entire perimeter of the ECR by boat. GPS points will be taken around the extent of the macrophyte bed to determine both its location and its extent (by area in square metre). Surveys will be conducted in the ECR from September to February as it is during the warmer months that emergent macrophytes will be growing and flowering. Location, extent (length / area covered) of

each emergent macrophyte species will be recorded. When the monitoring program commenced in 2010 it was anticipated that monthly sampling in spring and autumn would occur, but a decade on this sampling frequency may need to be revisited. Descriptive statistics (length, width and area covered) of each emergent macrophyte species will be calculated. In addition, a GIS based map showing locations of each macrophyte stand location will be derived.

RESULTS

Three stands of emergent macrophytes, all of Cumbungi (*Typha domingensis*), were found along the shoreline of the Cotter Reservoir in March and May 2022 (Figure 27). The first, located on the western shore just downstream of Bracks Hole road, measured approximately 2 x 2 m (4m²) in area (Figure 28). The other two were located next to each other near the old boat ramp road on the eastern shoreline of the reservoir (Figure 27). The largest of the two, measured approximately 9 x 4 m (36 m²) and formed the densest of the three stands (Table 14, Figure 29). The third comprised two loosely grouped sparse stands of approximately 2 x 2 m (4 m²) each (Table 14).

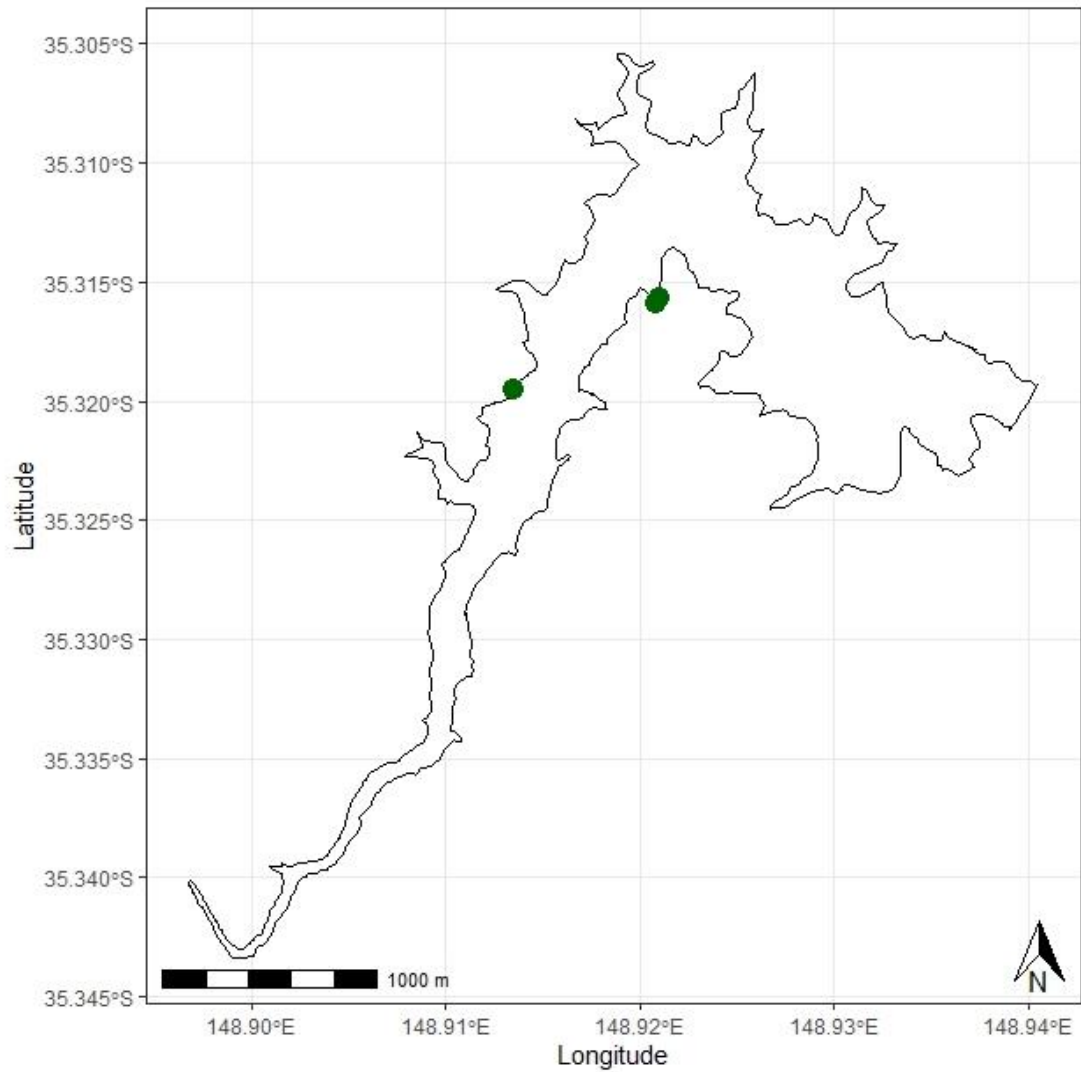


Figure 26. Map of the location of emergent macrophyte stands (green circles) along the shoreline of the Cotter Reservoir as detected in March and May 2022.

Table 14. Details of each emergent macrophyte stand in the Cotter Reservoir as of May 2022.

Species	Location	Approx. size (m ²)
Cumbungi (<i>Typha domingensis</i>)	Western shoreline, shoreline near Bracks Hole	4 m ²
Cumbungi (<i>Typha domingensis</i>)	Eastern shoreline, inlet near old boat ramp road	36 m ²
Cumbungi (<i>Typha domingensis</i>)	Eastern shoreline, inlet near old boat ramp road	2 x 4 m ²



Figure 27. Small stand of Cumbungi located on the western shoreline of Cotter Reservoir near Bracks Hole (photo taken 16/03/2022)



Figure 28. Largest stand of Cumbungi, in the inlet near the old boat ramp road on the eastern shoreline of Cotter Reservoir (photo taken 03/05/2022).

DISCUSSION AND CONCLUSIONS

For the first time since filling commenced in 2013, emergent macrophytes have been detected along the shoreline of the Cotter Reservoir. The three stands were all comprised of Cumbungi, which was a common emergent macrophyte in the Cotter Reservoir prior to enlargement (Roberts 2006). Establishment of emergent macrophytes, Cumbungi to date, in Cotter Reservoir is most likely attributable to very stable water level since September 2020 (Figure 3). Macrophytes provide a number of important services, including providing habitat and food for aquatic animals (e.g. fish, macroinvertebrates and birds) and stabilising sediments (Madsen *et al.* 2001, Richardson *et al.* 2002, Fleming *et al.* 2011). Emergent macrophytes were found to be crucially important refuge habitat for Macquarie perch in Cotter Reservoir prior to enlargement (Ebner and Lintermans 2007; Lintermans 2012). Whilst the current extent of emergent macrophytes in the Cotter Reservoir is very small (in comparison to other available habitats), and currently comprised of a single species it does indicate that stable water levels of the enlarged reservoir are suitable to macrophyte germination. The other formerly abundant emergent macrophyte Common Reed (*Phragmites australis*) has not yet re-established.

RECOMMENDATIONS

Based on the detection of emergent macrophytes in autumn 2022, it is recommended that formal surveys of emergent macrophytes commence. After discussions with Dr Will Higgsion (wetland vegetation ecologist, Centre for Applied Water Science, University of Canberra), we propose an alternation to the original proposed methods. We propose that monitoring of emergent macrophytes occur annually, during later summer early autumn. The reduction in frequency from the original methods and timing of the survey reflects;

- a) Emergent macrophyte extent is unlikely to alter detectably on the monthly scale
- b) Late summer / early autumn is the time at which the macrophyte stands will be at their maximum extent.

QUESTION 10: Are there adequate food resources (particularly decapods) for the Macquarie perch following the filling and operation of the enlarged Cotter Reservoir?

BACKGROUND

It was expected that as the ECR filled and became operational the food resources of Macquarie perch were likely to change. The substantial beds of emergent macrophytes that fringed the old Cotter Reservoir have been submerged, and the fluctuating water levels of an operational reservoir may prevent their reestablishment (Lintermans 2012). These reed beds supported significant densities of decapod crustaceans, particularly freshwater prawn (*Macrobrachium*) and shrimp (*Paratya*) which are favoured food items for Macquarie perch (Norris *et al.* 2012). During and following inundation a trophic upsurge was expected (and occurred) where food resources of Macquarie perch were plentiful and Macquarie perch had increased body condition (as experienced in other newly filled reservoirs such as Lake Dartmouth). As the reservoir ages, it was expected that the food resources of Macquarie perch may diminish and result in poorer body condition and this is likely to result in reduced fecundity and could lead to a negative impact on recruitment to the population.

METHODS

The sampling design follows that outlined in the Food Resources study of Norris *et al.* (2012) (the baseline samples) so comparisons with pre, during and post-filling can be made. All Macquarie perch food resources were targeted, with an emphasis on decapoda.

Table 15. Outline of the sampling design for Question 10 of the fish monitoring program.

Feature	Detail
Target species and life history phase	Food resources of Macquarie perch (primarily decapods).
Sampling technique/s	Edge sampling of each major habitat (3 each of rocky shore, bare shore, woody habitat and macrophyte – where possible) and plankton tows (1 in each longitudinal third of the reservoir).
Timing	Bi-annually in spring and autumn.
Number / location of sites	Conducted in the ECR only.
Information to be collected	Relative abundance and composition of food resources.
Data analysis	Relative abundance of prey items (with particular focus on decapods) was compared between phase (baseline, filling and operational), season and habitat type using three-way PERMANOVA and principal components analysis.

Food resources sampling was undertaken in autumn and spring in the ECR and followed the sampling and processing protocols of Norris *et al.* (2012). Each sampling event involved taking three replicate invertebrate samples of each habitat type occurring in the reservoir (of bare shore, rocky shore, timber and macrophyte when available). Sampling locations were determined by dividing the reservoir into three equal sections and sampling each habitat type per section. Invertebrate samples were collected from edge habitats with a sweep net (250 μm mesh) over a 10 m transect. Samples were then preserved in 70% ethanol for later processing in the laboratory. In the laboratory, samples were rinsed through a 250 μm mesh sieve to remove fine sediment and ethanol, and then placed in a large tray with water. Coarse scale invertebrate selection of entire edge habitat samples was performed using a magnifying lamp for one hour to calculate the numerical abundance of each invertebrate taxa. This method effectively captured information on the large quantities of abundant items such as decapods and generally resulted in the selection of larger invertebrates.

Fine scale invertebrate selection of 10% of the remaining sample under a stereomicroscope was then performed, and higher-powered magnification facilitated selection of smaller taxa. This was achieved by placing the remaining sample into a sub-sampler consisting of a box divided into 100 cells, 3 cm x 3 cm x 2.5 cm deep (Marchant 1989). The box was agitated until the sample was distributed evenly across cells. A total of 10 cells out of the 100 were randomly selected using two ten-sided dice, and their contents were removed with a vacuum pump. This standardised sampling method allowed for calculation of numerical abundance of taxa. All invertebrate identification was to order for aquatic taxa or a terrestrial item category for terrestrial-occurring invertebrates for edge habitat food availability analyses.

Plankton tows to collect invertebrates from open water habitats were also undertaken in three replicate sections of the reservoir (downstream, middle and upstream thirds). A weighted, modified 250 μm mesh net with a circular opening (300 mm wide) was lowered into the water column 50 m away from shore at 1 m depth and pulled by a two-person crew in motorised boat along a 50 m transect (distance was determined using a rangefinder) for each tow (sampling 3.54 m³ of open water habitat). Sampling was conducted bi-annually in spring and autumn at one site (the ECR). Open water invertebrates were identified from a 10% sub-sample using a 100 cell sub-sampler (Marchant 1989) as described for fine scale invertebrate selection of edge habitat samples above.

To analyse differences between the baseline samples of Norris *et al.* (2012) study and filling and operational edge samples, principal component analysis ordination and PERMANOVA analyses were conducted for each processing type (coarse pick and 10% subsample) as well as for the plankton tow samples. Principal Component analysis ordinations (PCO) of log₁₀ transformed data were arranged into resemblance matrices using the Bray-Curtis Similarity measure. Vectors are the raw Pearson's correlations for the taxa that are most ($r > 0.4$) correlated with each of the PCO axes. Unbalanced permutational analysis of variance (PERMANOVA) was conducted on coarse pick data and subsample data separately. Data was log₁₀ and a Bray-Curtis measure used for resemblance matrix. PERMANOVA analysis consisted of Phase, season and habitat as fixed factors. Highest interaction term was removed for repeated measures design. Type III Sum of Squares used to account for unbalanced (years across phase) design and the counts of each macroinvertebrate taxa used as variables. Decapod abundances derived from the coarse pick samples were analysed separately using ANOSIM with phase and season as factors.

RESULTS

Edge samples - Coarse pick

There was a significant difference in the coarse pick samples based on sampling phase, season and the phase x season interaction but no significant effect of habitat (Table 16). All phases were significantly different between season, except for between baseline and filling in spring, which was non-significant. PCO revealed that this difference between phases of coarse pick samples was largely driven by lower abundances of Coleoptera, Hemiptera and Diptera and higher abundances of Chironomids and terrestrial items in filling and operational phases compared to baseline monitoring (Figure 29). Decapod abundances were affected by season (Global R = 0.253, $p < 0.001$), with abundances being significantly higher in autumn compared to spring. Decapod abundances were not affected by phase (Global R = 0.024, $p = 0.247$). Decapod abundances in autumn have generally increased since 2018, after being low during early operational years (Figure 30). Decapod abundance in autumn 2022 was the second highest since monitoring began (Figure 30).

Table 16. Results of PERMANOVA analysis of coarse pick macroinvertebrate community composition in Cotter Reservoir from 2010 – 2022 (bold text indicates statistically significant difference at the 0.05 level).

Source	df	SS	MS	Pseudo-F	P(perm)	Unique permutations
Phase	2	25313	12656	9.5039	0.0001	9921
Season	1	6706.1	6706.1	5.0357	0.0004	9949
Habitat	2	965.36	482.68	0.36245	0.947	9928
Phase x Season	2	5669	2834.5	2.1285	0.0294	9930
Phase x Habitat	4	3110.7	777.68	0.58397	0.904	9926
Season x Habitat	2	1398.4	699.19	0.52503	0.8621	9930
Residuals	64	85230	1331.7			
Total	77	129790				

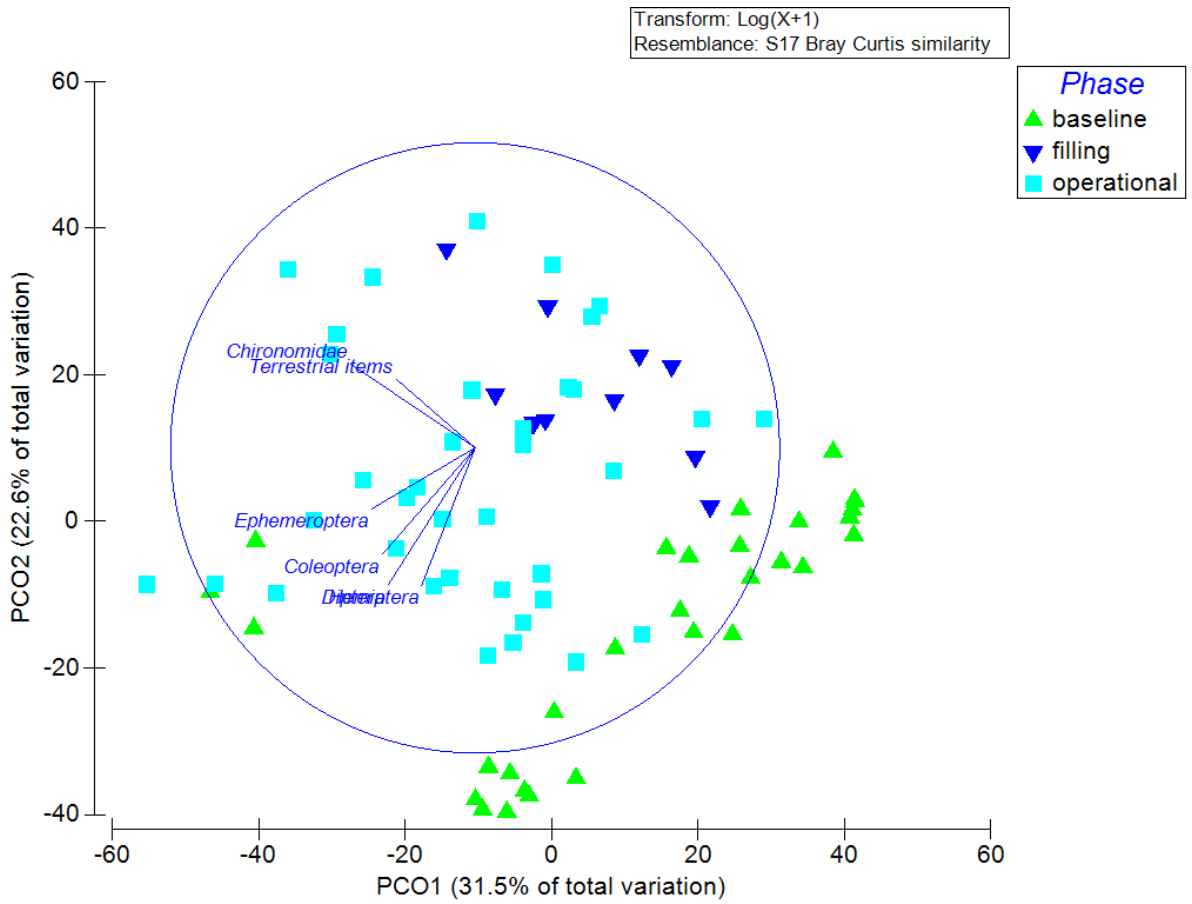


Figure 29. Graphical representation of a principal component analysis ordination of invertebrates from the coarse pick from spring and autumn monitoring in pre-filling (baseline) (data from Norris et al. 2012), filling phase (2013–2015) and operational phase (2016–2022).

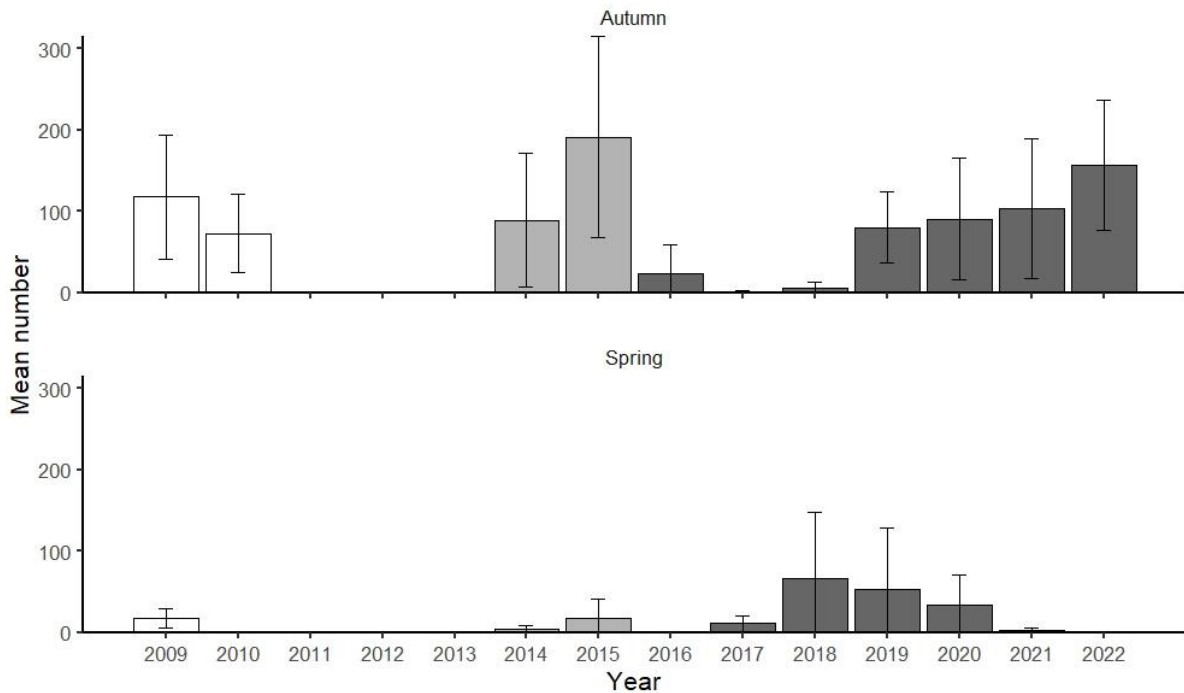


Figure 30. Relative abundance (mean \pm 95% confidence limits with Bonferroni corrections) of decapods collected from coarse pick edge samples taken from ECR during baseline (Pre-filling, 2009 / 2010; white bars) (Norris et al. 2012), filling (2013 – 2015; light grey bars) and operational (2016 – 2022; dark grey bars) monitoring periods for autumn and spring. Note: Spring 2022 has not been sampled at the time of reporting.

Edge samples - 10% sub-sample

As for the coarse pick, there was a significant difference in the 10% sub samples based on sampling phase and season, and a significant phase x season interaction (Table 17). All phases were significantly different between seasons except for between operational and filling in autumn, which was non-significant. All phases were significantly different from each other in the composition of the 10% subsamples taken from edge habitat. The difference between baseline and filling and operational phase 10% sub-samples in spring was largely driven by the higher abundances of Diptera and in the baseline samples (especially in 2010), and higher abundances of terrestrial items during filling and operational phases (Figure 31).

Table 17. Results of PERMANOVA analysis of 10% subsample macroinvertebrate community composition in Cotter Reservoir from 2009-2010 (baseline) & 2013 – 2015 (Filling) and 2016 – 2022 (Operational) (bold text indicates statistically significant difference at the 0.05 level).

Source	df	SS	MS	Pseudo-F	P(perm)	Unique permutations
Phase	2	26078	13039	9.1889	0.0001	9913
Season	1	10814	10814	7.6213	0.0001	9936
Habitat	2	2379.2	1189.6	0.83836	0.6391	9912
Phase x Season	2	15952	7976	5.621	0.0001	9903
Phase x Habitat	4	6216.5	1554.1	1.0952	0.3366	9878
Season x Habitat	2	1078.5	539.23	0.38001	0.9846	9917
Residuals	62	87976	1419			
Total	75	149420				

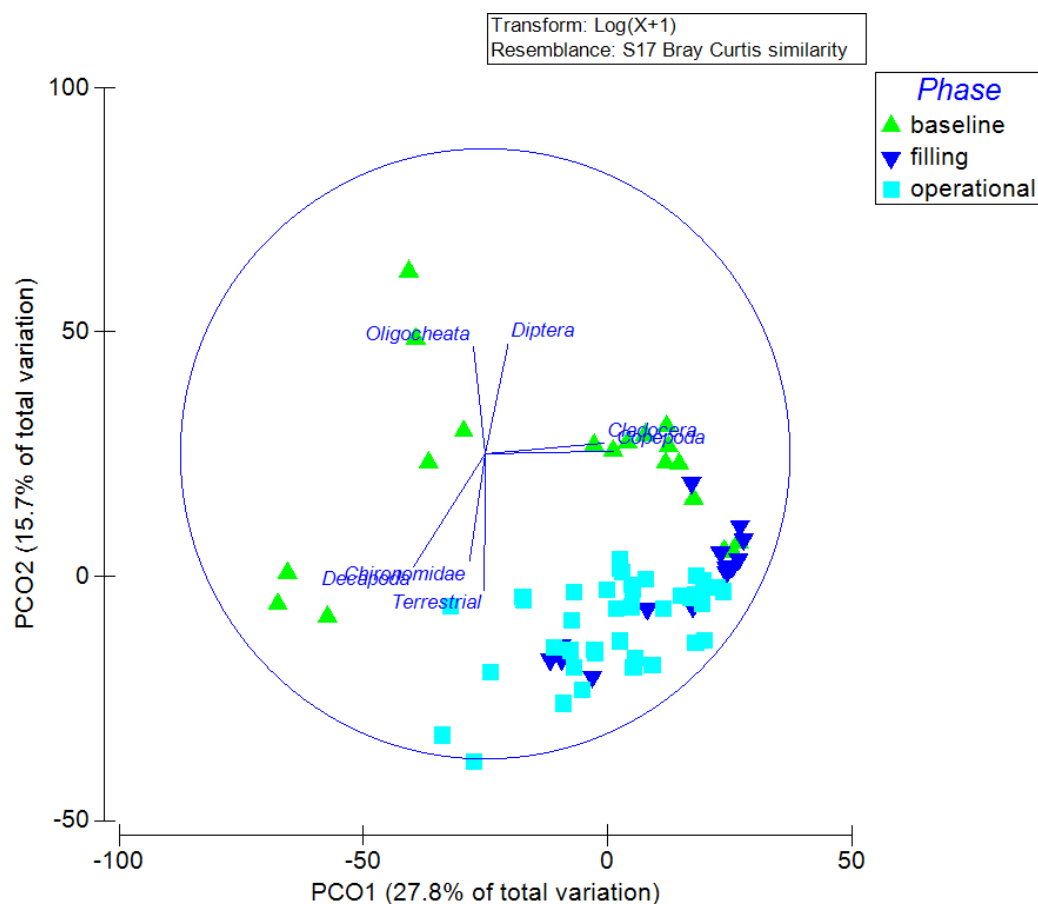


Figure 31. Graphical representation of a principal component analysis ordination of invertebrates from the 10% sub-sample processing from spring and autumn monitoring in baseline (data from Norris et al. 2012), filling phase (2013–2015) and operational phase (2016–2022).

Tow samples

Microcrustaceans dominated the plankton tow samples from the baseline and the filling and operational phase monitoring, comprising 99.9%, 100% and 100% of samples, respectively. Although there are some interannual variations in the microcrustacean community, there was no significant effect of phase or season (Table 18 & Figure 32).

Table 18. Results of PERMANOVA analysis of tow sample microcrustacean community composition in Cotter Reservoir from 2009-2010 (baseline) & 2013 – 2015 (Filling) and 2016 – 2022 (Operational) (bold text indicates statistically significant difference at the 0.05 level).

Source	df	SS	MS	Pseudo-F	P(perm)	Unique permutations
Phase	2	338.59	169.29	0.93249	0.3657	9927
Season	1	185.26	185.26	1.0204	0.2916	9933
Phase x Season	2	259.06	129.53	0.71346	0.481	9941
Residuals	51	9259.1	181.55			
Total	56	9754.4				

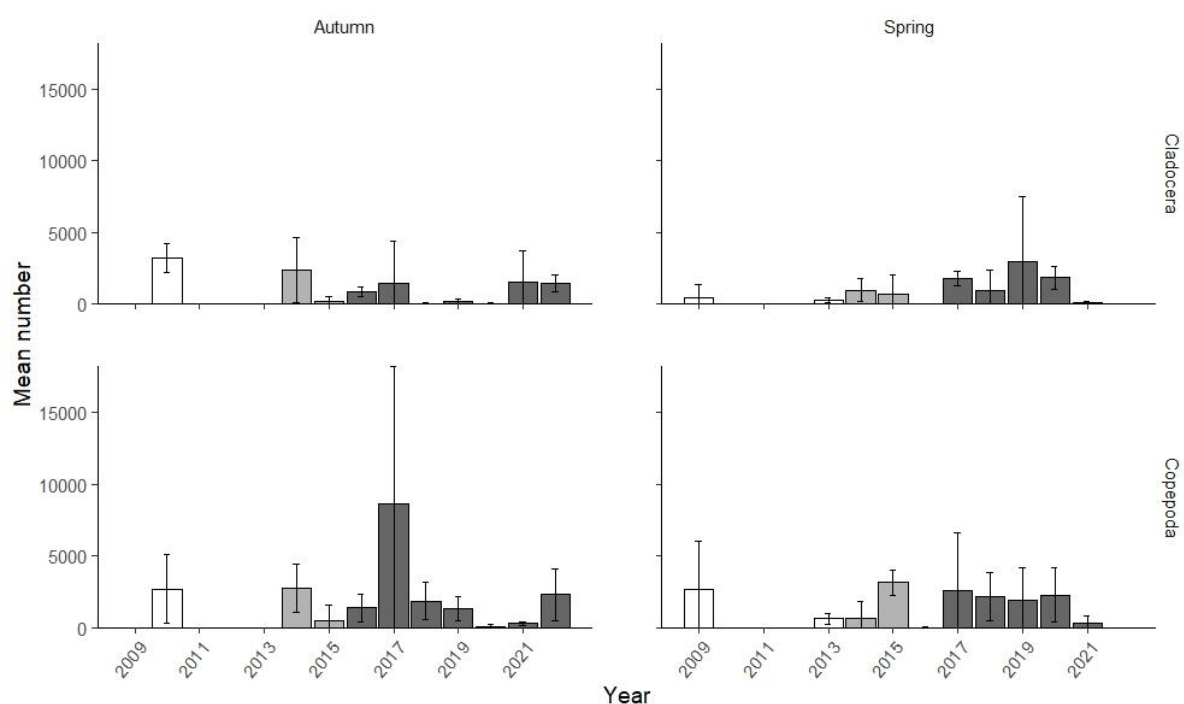


Figure 32. Relative abundance (mean \pm 95% confidence limits with Bonferroni corrections) of each microcrustacean taxa collected in autumn and spring of each phase of monitoring phase in ECR during baseline (Pre-filling, 2009 / 2010; white bars) (Norris et al. 2012), filling (2013 – 2015; light grey bars) and operational (2016 – 2022; dark grey bars) monitoring periods. Note: Spring 2022 had not been collected at time of reporting.

DISCUSSION AND CONCLUSIONS

Decapod abundances in coarse pick edge samples are similar between phases. Decapod abundance in autumn during early operational phase monitoring was lower than that observed for autumn in baseline, filling and latter operational phase monitoring. Decapod abundance was much lower in spring than autumn. Despite being in very low abundances in early years of the operational phase, decapod abundance has been generally increasing since spring 2018, with decapod abundances now similar to that of baseline and filling phase monitoring (see Figure 30). Decapods have been previously found to be an important food item of adult Macquarie perch and may be an important antecedent factor in spawning success as previous studies have found Macquarie perch fecundity to be positively related to body condition (Gray *et al.* 2000, Lintermans 2006, Norris *et al.* 2012, Hatton 2016). Decapod abundances have stabilised in the latter years of operational phase, and it appears as though the operation of Cotter Reservoir is having minimal impact since 2018 on this important food resource of Macquarie perch.

Terrestrial items were more abundant in the coarse pick samples in the filling and operational phase monitoring compared to the pre-filling/baseline study. As terrestrial habitats (earth and vegetation) become inundated, terrestrial invertebrates will enter the water column. Also, whilst the reservoir is full, overhanging vegetation would provide a source of terrestrial insects to the reservoir. This would explain the increased abundance of terrestrial items in the filling and operational phase monitoring. Macquarie perch are an opportunistic feeder in Cotter Reservoir (Norris *et al.* 2012), and it is likely that they will take advantage of terrestrial items present during filling (Cadwallader and Douglas 1986). Indeed, data from stomach flushing showed that Macquarie perch were feeding on earthworms during spring 2013 in the Cotter Reservoir, but that this dietary item was not important in the following year (Hatton 2016).

Tow net samples were numerically dominated by the microcrustaceans Cladocera and Copepoda in all phases. Abundances of both taxa varied, but there was no significant effect of phase or season on abundances. Cladocera were found to be an important dietary item of Macquarie perch in the Cotter Reservoir prior to filling (Norris *et al.* 2012) and also in another reservoir study of Macquarie perch in Lake Dartmouth (Cadwallader and Douglas 1986), but had a reduced importance whilst the reservoir was filling (Hatton 2016). Population abundance of Cladocera and Copepods are largely driven by temperature, turbidity, water residence time and predation (Dejen *et al.* 2004, Obertegger *et al.* 2007, Silva *et al.* 2014, Bartrons *et al.* 2015). Effect of the operation of Cotter Reservoir on microcrustacean abundances at this stage appears to be minimal.

RECOMMENDATIONS

Sampling for this question follows previously developed methods and appears to be adequate for detecting change. No change the monitoring approach is recommended.

The majority of the food resource differences between phases likely fall within natural annual and sampling variation. The main change of importance to the resident Macquarie perch population is the reduction in decapod abundance during early operational phase, though this appears to be increasing in latter operational years. So far, this has not appeared to have a negative effect on adult

condition, spawning or survival and growth or juveniles. No management intervention is recommended. It is now eight years since diet of Macquarie perch in the ECR was investigated, and Macquarie perch diet has not been assessed since the Cotter Reservoir has entered operational phase. A re-examination of Macquarie perch diet is recommended.

REFERENCES

- ACTEW Corporation. 2009a. *Enlargement of Cotter Reservoir and associated works. Final public environment report 2008/4524*. ACTEW Corporation, Canberra.
- ACTEW Corporation. 2009b. *Enlargement of the Cotter Reservoir and associated works. Environmental Impact Statement*. ACTEW Corporation, Canberra.
- ACTEW Corporation. 2013. *Alien fish management plan: Enlarged Cotter Dam*. ACTEW Corporation, Canberra.
- Anderson, M. J., R. N. Gorley, and K. R. Clarke. 2008. *PERMANOVA+ for PRIMER: Guide to Software and Statistical Methods*. PRIMER-E, Plymouth, UK.
- Bartrons, M., Á. Einarsson, R. L. Nobre, C. M. Herren, K. C. Webert, S. Brucet, S. R. Ólafsdóttir, and A. R. Ives. 2015. Spatial patterns reveal strong abiotic and biotic drivers of zooplankton community composition in Lake Mývatn, Iceland. *Ecosphere* **6**:1-20.
- Blanchet, S., G. Loot, and J. Dodson. 2008. Competition, predation and flow rate as mediators of direct and indirect effects in a stream food chain. *Oecologia* **157**:93-104.
- Broadhurst, B., R. Clear, and M. Lintermans. 2016a. *Potential barriers to upstream migration of Macquarie perch to spawning areas in the Cotter River*, ACT Institute for Applied Ecology, University of Canberra, Canberra.
- Broadhurst, B., R. C. Clear, C. Fulton, and M. Lintermans. 2018. *Enlarged Cotter Reservoir ecological monitoring program: technical report 2018*. Report, Institute for Applied Ecology, University of Canberra, Canberra.
- Broadhurst, B., R. C. Clear, C. Fulton, and M. Lintermans. 2019. *Enlarged Cotter Reservoir ecological monitoring program: technical report 2019*. Report, Institute for Applied Ecology, University of Canberra, Canberra.
- Broadhurst, B., J. Dyer, B. Ebner, J. Thiem, and P. Pridmore. 2011. Response of two-spined blackfish *Gadopsis bispinosus* to short-term flow fluctuations in an upland Australian stream. *Hydrobiologia* **673**:63-77.
- Broadhurst, B., B. Ebner, and R. Clear. 2012a. A rock-ramp fishway expands nursery grounds of the endangered Macquarie perch *Macquaria australasica*. *Australian Journal of Zoology* **60**:91–100.
- Broadhurst, B. T., R. C. Clear, C. Fulton, H. Allan, and M. Lintermans. 2020. *Enlarged Cotter Reservoir ecological monitoring program: technical report 2020.*, Institute for Applied Ecology, University of Canberra, Canberra.
- Broadhurst, B. T., R. C. Clear, C. Fulton, H. Allan, and M. Lintermans. 2021. *Enlarged Cotter Reservoir ecological monitoring program: technical report 2021.*, Institute for Applied Ecology, University of Canberra, Canberra.
- Broadhurst, B. T., R. C. Clear, C. Fulton, and M. Lintermans. 2015. *Enlarged Cotter Reservoir ecological monitoring program: technical report 2015*. Institute for Applied Ecology, University of Canberra, Canberra.
- Broadhurst, B. T., R. C. Clear, C. Fulton, and M. Lintermans. 2016b. *Enlarged Cotter Reservoir ecological monitoring program: technical report 2016*. Institute for Applied Ecology, University of Canberra, Canberra.
- Broadhurst, B. T., R. C. Clear, C. Fulton, and M. Lintermans. 2017. *Enlarged Cotter Reservoir ecological monitoring program: technical report 2017*. Institute for Applied Ecology, University of Canberra, Canberra.

- Broadhurst, B. T., R. C. Clear, M. Lintermans, and C. Fulton. 2014. *Enlarged Cotter Reservoir ecological monitoring program: technical report 2014*. Institute for Applied Ecology, University of Canberra, Canberra.
- Broadhurst, B. T., M. Lintermans, J. D. Thiem, B. C. Ebner, D. W. Wright, and R. C. Clear. 2012b. Spatial ecology and habitat use of two-spined blackfish *Gadopsis bispinosus* in an upland reservoir. *Aquatic Ecology* **46**:297–309.
- Budy, P., G. P. Thiede, J. Lobón-Cerviá, G. G. Fernandez, P. McHugh, A. McIntosh, L. A. Vøllestad, E. Becares, and P. Jellyman. 2013. Limitation and facilitation of one of the world's most invasive fish: an intercontinental comparison. *Ecology* **94**:356-367.
- Cadwallader, P. L., and J. D. Douglas. 1986. Changing food habits of Macquarie perch *Macquaria australasica* Cuvier (Pisces: Percichthyidae). during the initial filling phase of Lake Darmouth, Victoria. *Australian Journal of Marine and Freshwater Research* **37**:647–657.
- Dejen, E., J. Vijverberg, L. A. Nagelkerke, and F. A. Sibbing. 2004. Temporal and spatial distribution of microcrustacean zooplankton in relation to turbidity and other environmental factors in a large tropical lake (L. Tana, Ethiopia). *Hydrobiologia* **513**:39-49.
- Dorfman, E. J., and M. J. Kingsford. 2001a. Environmental determinants of distribution and foraging behaviour of cormorants (*Phalacrocorax* spp.) in temperate estuarine habitats. *Marine Biology* **138**:1–10.
- Dorfman, E. J., and R. T. Kingsford. 2001b. Scale-dependent patterns of abundance and habitat use by cormorants in and Australia and the importance of nomadism. *Journal of Arid Environments* **49**:677-694.
- Ebner, B., B. Broadhurst, M. Lintermans, and M. Jekabsons. 2007. A possible false negative: lack of evidence for trout predation on a remnant population of the endangered Macquarie perch, *Macquaria australasica*, in Cotter Reservoir, Australia. *New Zealand Journal of Marine and Freshwater Research* **41**:231–237.
- Ebner, B., and M. Lintermans. 2007. *Fish passage, movement requirements and habitat use for Macquarie perch. Final report to the Department of Agriculture, Fisheries and Forestry Australia*. Parks, Conservation and Lands, Canberra.
- Ebner, B., J. Thiem, B. Broadhurst, R. Clear, and K. Frawley. 2008. *Delivering environmental flows to large biota. Final Report to the Department of the Environment, Water, Heritage and the Arts*. Parks, Conservation and Lands, Canberra.
- Farrington, L. W., M. Lintermans, and B. C. Ebner. 2014. Characterising genetic diversity and effective population size in one reservoir and two riverine populations of the threatened Macquarie perch. *Conservation Genetics*:1-10.
- Fleming, J. P., J. D. Madsen, and E. D. Dibble. 2011. Macrophyte re-establishment for fish habitat in Little Bear Creek Reservoir, Alabama, USA. *Journal of Freshwater Ecology* **26**:105-114.
- Gray, S. C., S. S. De Silva, B. A. Ingram, and G. J. Gooley. 2000. Effects of river impoundment on body condition and reproductive performance of the Australian native fish, Macquarie perch (*Macquaria australasica*). *Lakes & Reservoirs: Research and Management* **5**:281–291.
- Hatton, S. 2016. *Ecological responses to the initial filling of an enlarged temperate reservoir: Cotter Reservoir (ACT, Australia)*. PhD thesis. University of Canberra, Canberra.
- Icon Water Limited. 2019. *Cotter Reservoir Fish Management Plan (FMP). Version 4*. Icon Water Limited, Canberra.

- Jonsson, N., T. Næsje, B. Jonsson, R. Saksgård, and O. Sandlund. 1999. The influence of piscivory on life history traits of brown trout. *Journal of Fish Biology* **55**:1129-1141.
- Kimmel, B. L., and A. W. Groeger. 1986. Limnological and ecological changes associated with reservoir aging. in G. E. Hall and M. J. Van Den Avyle, editors. *Reservoir Fisheries Management: Strategies for the 80's*. Reservoir Committee, South Division American Fisheries Society, Bethesda, Maryland.
- Kingsford, R. T., A. L. Curtin, and J. L. Porter. 1999. Water flows on Cooper Creek in arid Australia determine 'boom' and 'bust' periods for waterbirds. *Biological Conservation* **88**:231–248.
- Lintermans, M. 1998. *The ecology of the two-spined blackfish Gadopsis bispinosus (Pisces: Gadopsidae)*. Unpublished M. Sc. Thesis. M Sc Thesis, School of Botany and Zoology, Australian National University, Canberra.
- Lintermans, M. 2001. *Fish Monitoring Program to Assess the Effectiveness of Environmental Flows in the Cotter and Queanbeyan rivers*. Consultancy, Ecowise Environmental Ltd, Canberra.
- Lintermans, M. 2002. *Fish in the Upper Murrumbidgee Catchment: A Review of Current Knowledge*. Environment ACT, Canberra.
- Lintermans, M. 2005a. *ACT future water options fish impact study: a review of potential impacts on fish and crayfish of future water supply options for the Australian Capital Territory: stage 1*. ACTEW Corporation, Canberra.
- Lintermans, M. 2005b. *Environmental flows in the Cotter River, ACT, and the response on the threatened fish species Macquaria australasica and Gadopsis bispinosus in 2003 and 2004*. Environment ACT, Canberra.
- Lintermans, M. 2006. *The re-establishment of endangered Macquarie perch Macquaria australasica in the Queanbeyan River, New South Wales, with an examination of dietary overlap with alien trout*. Cooperative Research Centre for Freshwater Ecology, Canberra.
- Lintermans, M. 2007. *Fishes of the Murray-Darling Basin: an introductory guide*. Murray-Darling Basin Commission, Canberra.
- Lintermans, M. 2012. Managing potential impacts of reservoir enlargement on threatened *Macquaria australasica* and *Gadopsis bispinosus* in southeastern Australia. *Endangered Species Research* **16**:1–16.
- Lintermans, M. 2013. *Using translocation to establish new populations of Macquarie perch, Trout cod and Two-spined blackfish in the Canberra region. Final report to ACTEW Water*. Institute for Applied Ecology, University of Canberra, Canberra.
- Lintermans, M. 2016. Finding the needle in the haystack: comparing sampling methods for detecting an endangered freshwater fish. *Marine and Freshwater Research* **67**:1740-1749.
- Lintermans, M. 2017. *Research into the establishment of new populations of Macquarie perch, through translocation: Final Report*. Institute for Applied Ecology, University of Canberra, Canberra.
- Lintermans, M., B. Broadhurst, and R. Clear. 2011. *Characterising potential predation of Macquarie perch Macquaria australasica by cormorants in Cotter Reservoir*. Institute for Applied Ecology, Canberra.
- Lintermans, M., B. Broadhurst, and R. Clear. 2013. *Assessment of the potential impacts on threatened fish from the construction, filling and operation of the Enlarged Cotter*

- Dam Phase 1 (2010-2012): Final Report. Report to ACTEW Corporation.* Institute for Applied Ecology, University of Canberra, Canberra.
- Lintermans, M., B. Broadhurst, J. Thiem, B. Ebner, D. Wright, R. Clear, and R. Norris. 2010. *Constructed homes for threatened fishes in the Cotter River catchment: Phase 2 final report. Report to ACTEW Corporation.* Institute for Applied Ecology, University of Canberra, Canberra.
- Macdonald, J., and Z. Tonkin. 2008. A review of the impacts of eastern gambusia on native fishes of the Murray–Darling Basin. *Murray–Darling Basin Authority Publication.*
- Madsen, J. D., P. A. Chambers, W. F. James, E. W. Koch, and D. F. Westlake. 2001. The interaction between water movement, sediment dynamics and submersed macrophytes. *Hydrobiologia* **444**:71-84.
- Marchant, R. 1989. A sub-sampler for samples of benthic macroinvertebrates. . *Bulletin of the Australian Society for Limnology* **12**:49–52.
- Merrick, J. R., and G. E. Schmida. 1984. *Australian freshwater fishes: biology and management.* J. Merrick, North Ryde, NSW.
- Miller, B. 1979. Ecology of the little black cormorant, *Phalacrocorax sulcirostris*, and the little pied cormorant, *P. melanoleucos*, in inland New South Wales. I. Food and feeding habits. *Australian Wildlife Research* **6**:79–95.
- Norris, R., D. Wright, M. Lintermans, D. Bourke, and E. Harrison. 2012. *Food resources for Macquarie perch in Cotter Reservoir. Final report to the Bulk Water Alliance.* Institute for Applied Ecology, University of Canberra., Canberra.
- NSW Fisheries. 2003. *Environmental Impact Statement on Freshwater Fish Stocking in NSW.* NSW Fisheries, Cronulla.
- O'Brien, W. J. 1990. Chapter 8. Perspectives on fish in reservoir ecosystems. Pages 209–226. *in* K. W. Thornton, B. L. Kimmel, and F. E. Payne, editors. *Reservoir limnology: Ecological perspectives.* John Wiley & Sons Inc., New York.
- Obertegger, U., G. Flaim, M. G. Braioni, R. Sommaruga, F. Corradini, and A. Borsato. 2007. Water residence time as a driving force of zooplankton structure and succession. *Aquatic Sciences* **69**:575-583.
- Ploskey, G. R. 1986. Effects of water-level changes on reservoir ecosystems, with implications for fisheries management. *in* G. E. Hall and M. J. Van Den Avyle, editors. *Reservoir fisheries management: strategies for the 80's.* . Reservoir Committee, Southern Division American Fisheries Society., Bethesda, Maryland.
- Pyke, G. H. 2005. A Review of the Biology of *Gambusia affinis* and *G. holbrooki*. *Reviews in Fish Biology and Fisheries* **15**:339-365.
- Richardson, S. M., J. M. Hanson, and A. Locke. 2002. Effects of impoundment and water-level fluctuations on macrophyte and macroinvertebrate communities of a dammed tidal river. *Aquatic Ecology* **36**:493-510.
- Ricker, W. E. 1975. Computation and interpretation of biological statistics of fish populations. *Bulletin of the Fisheries Research Board of Canada* **191**:1-382.
- Roberts, J. 2006. *Wetland Vegetation around Cotter Reservoir.* Canberra, ACT.
- Robinson, W. 2009. *Biometrical review of proposed fish monitoring programs for the Enlarged Cotter Dam project.* University of the Sunshine Coast.
- Ropert-Coudert, Y., D. Gremillet, and A. Kato. 2006. Swim speeds of free-ranging great cormorants. *Marine Biology* **149**:415-422.

- Ryan, K. A. 2010. *Macquarie perch under pressure: predation risk to an endangered fish population*. Unpublished PhD thesis. Institute for Applied Ecology, University of Canberra, Canberra.
- Ryan, K. A., M. Lintermans, B. C. Ebner, and R. Norris. 2013. Using fine-scale overlap in predator-prey distribution to assess avian predation risk to a reservoir population of threatened Macquarie Perch. *Freshwater Science* **32**:1057-1072.
- Silva, L. H., V. L. Huszar, M. M. Marinho, L. M. Rangel, J. Brasil, C. D. Domingues, C. C. Branco, and F. Roland. 2014. Drivers of phytoplankton, bacterioplankton, and zooplankton carbon biomass in tropical hydroelectric reservoirs. *Limnological-Ecology and Management of Inland Waters* **48**:1-10.
- Todd, C. R., and M. Lintermans. 2015. Who do you move: a stochastic population model to guide translocation strategies for an endangered freshwater fish in southeastern Australia. *Ecological Modelling* **311**:63–72.
- Todd, C. R., M. Lintermans, S. Raymond, and J. Ryall. 2017. Assessing the impacts of reservoir expansion using a population model for a threatened riverine fish. *Ecological Indicators* **80**:204-214.
- Tonkin, Z., J. Kearns, J. Lyon, S. R. Balcombe, A. J. King, and N. R. Bond. 2017. Regional-scale extremes in river discharge and localised spawning stock abundance influence recruitment dynamics of a threatened freshwater fish. *Ecohydrology* **10**:e1842.
- van Dijk, A. I., H. E. Beck, R. S. Crosbie, R. A. de Jeu, Y. Y. Liu, G. M. Podger, B. Timbal, and N. R. Viney. 2013. The Millennium Drought in southeast Australia (2001–2009): Natural and human causes and implications for water resources, ecosystems, economy, and society. *Water Resources Research* **49**:1040-1057.
- Winemiller, K. O., A. S. Flecker, and D. J. Hoeinghaus. 2010. Patch dynamics and environmental heterogeneity in lotic ecosystems. *Journal of the North American Benthological Society* **29**:84-99.